



**All for one and one for all:
the true potential of
whole-genome sequencing.**

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Center for Food Safety and Applied Nutrition,
US Food and Drug Administration

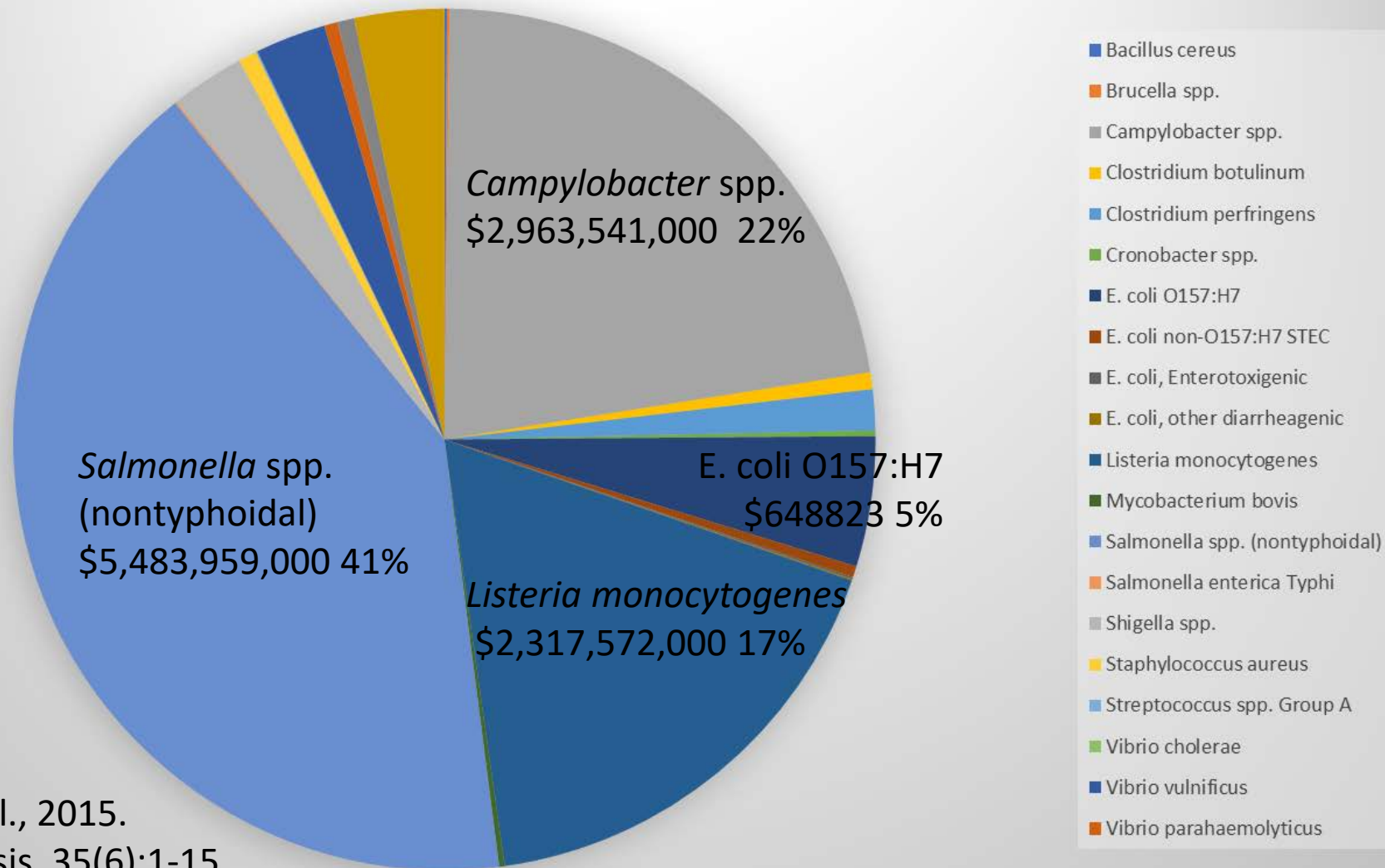
GLOBAL WATER FOOD SAFETY SUMMIT
College Park, MD, November 2019

Economic Analysis of *Salmonella* outbreaks annual reduction using WGS

	Canada	United States
Incidence of illness	47,028	1,200,000
Costs to adopt WGS	\$158,340,000	\$100,000,000
QUALY lost	469.75	16,782
Total Illness costs	\$287,770,000	\$3,300,000,000
Total net benefit of adopting WGS	\$90,250,000	\$1,000,000,000

Model assumes up to 70% reduction in numbers of illnesses due to WGS implementation. Benefits gained due to earlier detection and decreased time to recall food items. United States estimates are adjusted based on increase population size. Additional analysis is needed to adjust to US illnesses and US health care costs.

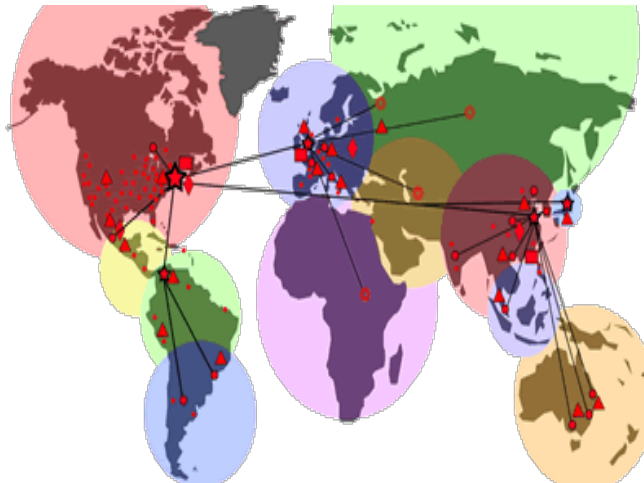
Monetary Loss from Bacteria Foodborne Illness Total \$13,279,603,000



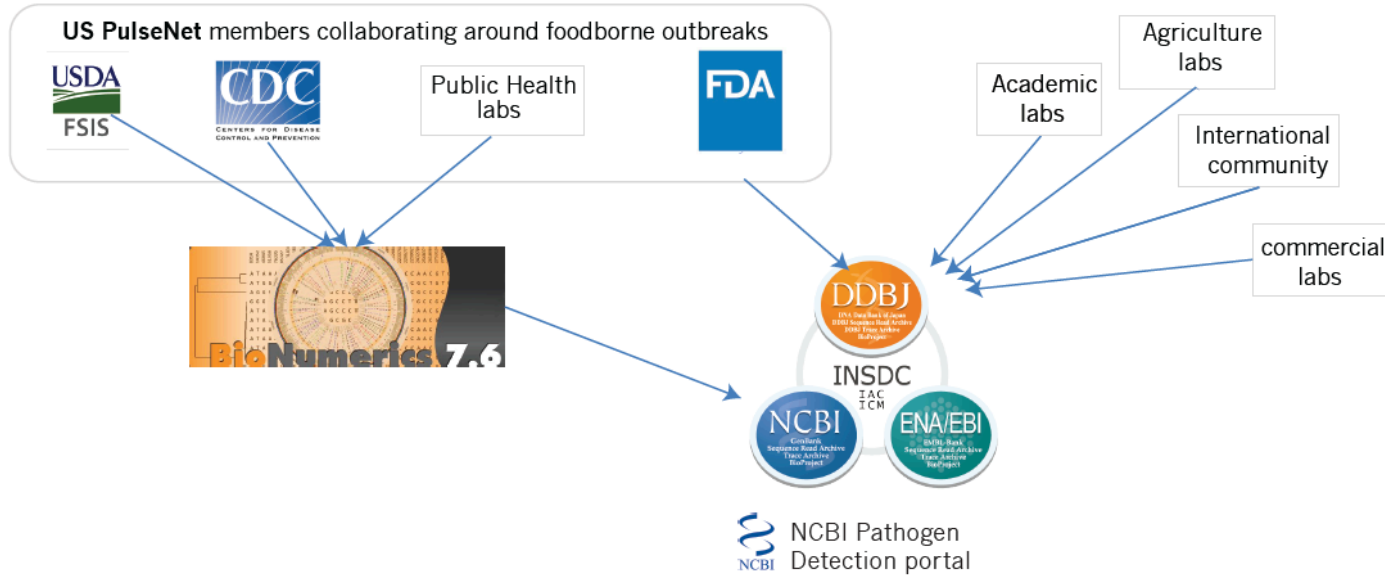
Minor et al., 2015.
Risk Analysis. 35(6):1-15

Why Develop a WGS based Network?

- Tracking and Tracing of food pathogens
 - Insufficient resolution of current tools
 - matching clinical to environmental
 - improve the environmental database
 - Faster identification of the food involved in the outbreak
 - Limited number of investigators vs. facilities and import lines
 - Global travel
 - Global food supply



Independent submission model:



Best practices document, Sept. 2019:

1. Check the quality of your entire run:

GalaxyTrakr, MicroRunQC workflow:



QC report for entire MiSeq run:

File	1	2	3	4	5	6	7	8	9	10	11	12
	Contigs	Length	GC%	N50	MedianInsert	MeanLength_R1	MeanLength_R2	MeanQ_R1	MeanQ_R2	Sequencing	13	14
FDAM000118_118_13011.fasta	99	4888884	58.3	157759	222	150.5	151.4	35.3	34.5	sentinella	19	19
FDAM000411_118_13011.fasta	113	4625033	51.3	64830	256	207.7	206.6	35.7	35.5	sentinella	4	4
FDAM011847_118_13011.fasta	76	4738332	50.78	244493	195	184.8	183.7	35.5	35.2	sentinella	19	19
FDAM011401_118_13011.fasta	46	4665022	57.1	324499	345	234.8	234.9	35.3	35.3	sentinella	23	23
FDAM011406_118_13011.fasta	77	5021371	51.5	173949	270	217.9	219.3	35.7	35.4	sentinella	19	19
FDAM011407_118_13011.fasta	80	4756952	53.0	182254	212	195.2	194.7	35.8	35.8	sentinella	19	19
FDAM011408_118_13011.fasta	82	4706024	53.14	175612	248	220.8	226.5	35.3	35.4	sentinella	19	19
FDAM011409_118_13011.fasta	80	4806251	57.6	182376	219	199.1	199.8	35.8	35.8	sentinella	19	19
FDAM011410_118_13011.fasta	83	4790239	55.9	182376	304	235.0	226.2	35.7	35.5	sentinella	19	19
FDAM011411_118_13011.fasta	82	4790214	55.9	182376	200	197.8	188.1	35.7	35.1	sentinella	19	19

assembly quality and coverage library quality Read quality sequence type

2. Detailed protocol for direct NCBI submissions:

Utilizing the Public GenomeTrakr Database for Foodborne Pathogen Traceback

Ruth E. Timme, Maria Sanchez Leon, and Marc W. Allard

Abstract

This protocol outlines the all the steps necessary to become a GenomeTrakr data contributor. GenomeTrakr is an international genomic reference database of mostly food and environmental isolates from foodborne pathogens. The data and analyses are housed at the National Center for Biotechnology Information (NCBI), which is a database freely available to anyone in the world. The Pathogen Detection browser at NCBI computes daily cluster results adding the newly submitted data to the existing phylogenetic clusters of closely related genomes. Contributors to this database can see how their new isolates are related to the real-time foodborne pathogen surveillance program established in the USA and a few other countries, and at the same time adding valuable new data to the reference database.

Key words: GenomeTrakr, WGS, Surveillance, Genomics, Foodborne pathogens, Traceback

1 Introduction

1.1 Inception of GenomeTrakr Within the FDA Mission

In 2012 FDA began a pilot project called GenomeTrakr to build a public genomic reference database of historical food and environmental isolates of *Salmonella*. The goal of this project was to improve the accuracy and response time for identifying the causes of foodborne outbreaks, to identify harborage in facilities, and to aid in establishing preventative controls [1]. In this pilot WGS data were collected by a distributed set of public health laboratories, transferred to the FDA for quality screening, then uploaded under an umbrella BioProject at NCBI's SRA database (Fig. 1). The result has been a continuously growing database of genomic sequence information and accompanying metadata (e.g., geographic loca-

3. Data curation and data management recommendations:

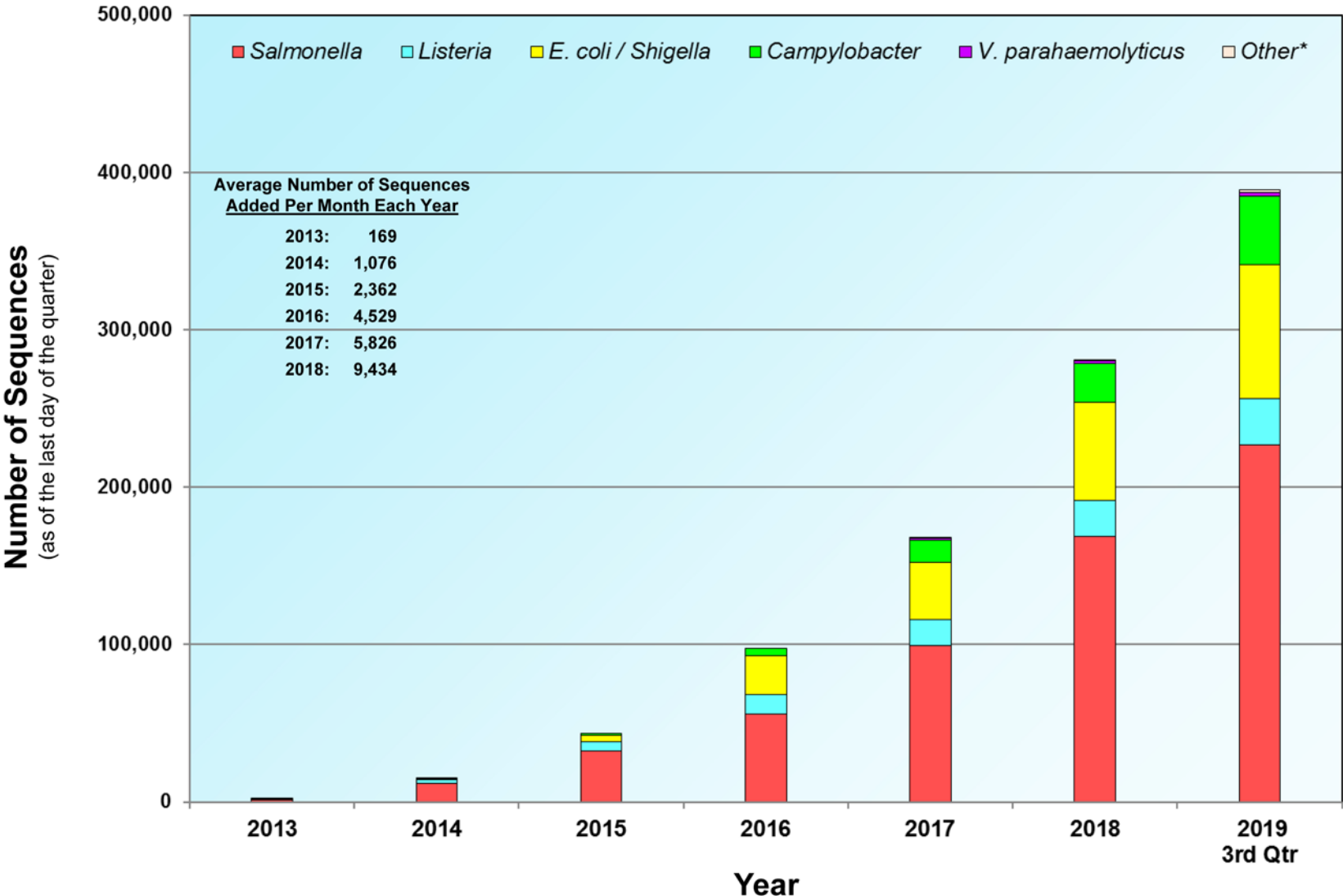
- Importance of data curation.
- Where are my data? How do I update/retract records in the following NCBI databases?
 - BioProject
 - BioSample
 - SRA
 - GenBank
 - NCBI Pathogen Detection portal
- Recommendations for managing your data, submissions, and accessions.

References:

Timme RE, Sanchez Leon M, Allard MW 2019. Utilizing the Public GenomeTrakr Database for Foodborne Pathogen Traceback. *Methods in molecular biology* (Clifton, N.J.) 1918:201–212.

Daniel Benisatto et al. In review. GalaxyTrakr: A Distributed Analysis Tool for Public Health Whole Genome Sequence Data Accessible to Non-Bioinformaticans.

Total Number of Sequences in the GenomeTrakr Database



First sequences uploaded in February 2013

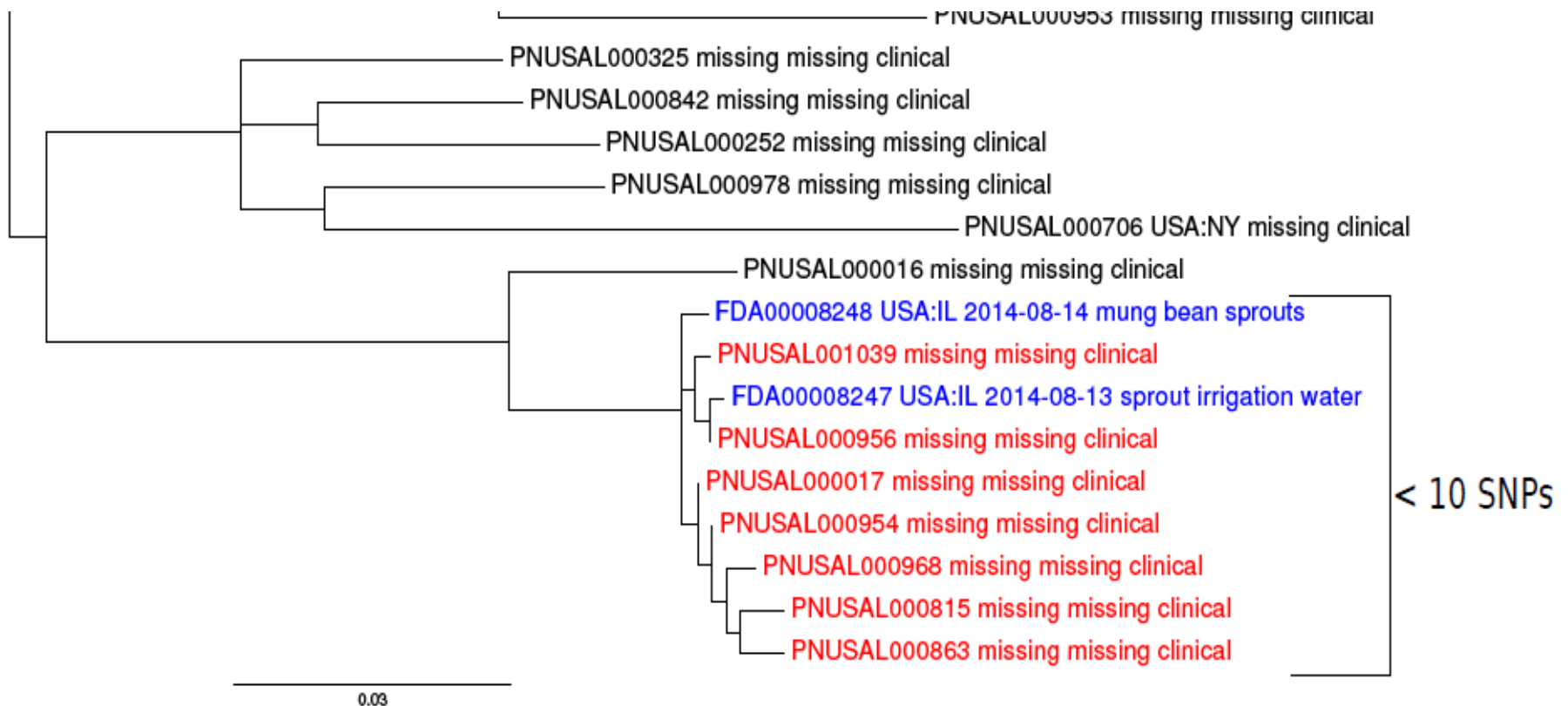
* Other pathogens: *Cronobacter*, *V. vulnificus*, *C. botulinum*, and *C. perfringens*

September 30, 2019 GenomeTrakr Numbers



Species	Total Isolates
<i>Salmonella enterica</i>	277,051
<i>E.coli and Shigella</i>	85,157
<i>Campylobacter jejuni</i>	43,705
<i>Listeria monocytogenes</i>	28,978
<i>Vibrio parahaemolyticus</i>	2,519
<i>Cronobacter</i>	636
<i>Vibrio vulnificus</i>	359
<i>Clostridium botulinum</i>	298
<i>Clostridium perfringens</i>	242
Total	388,945

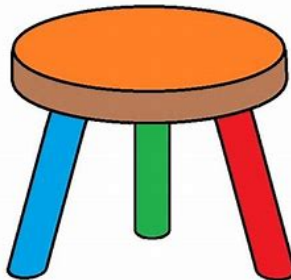
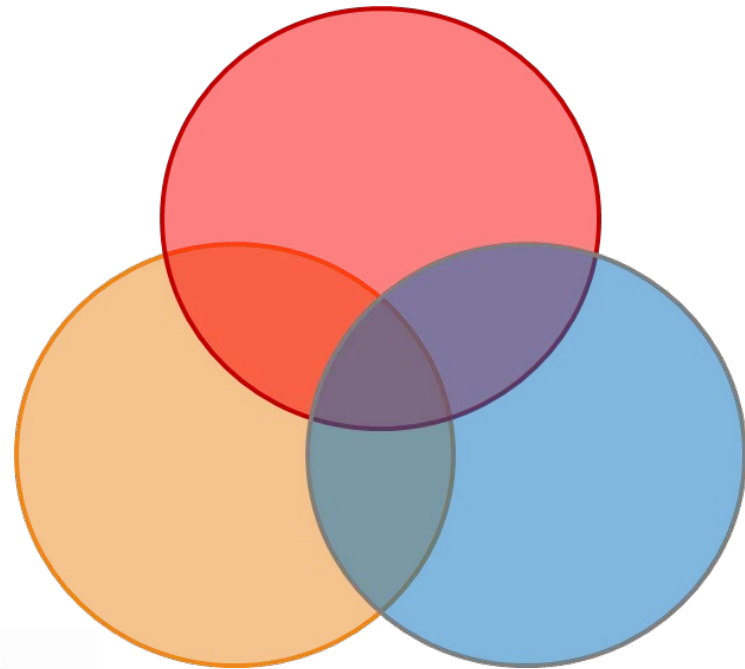
How do we use the GenomeTrakr information? Example of Listeria in sprouts using a phylogenetic perspective.

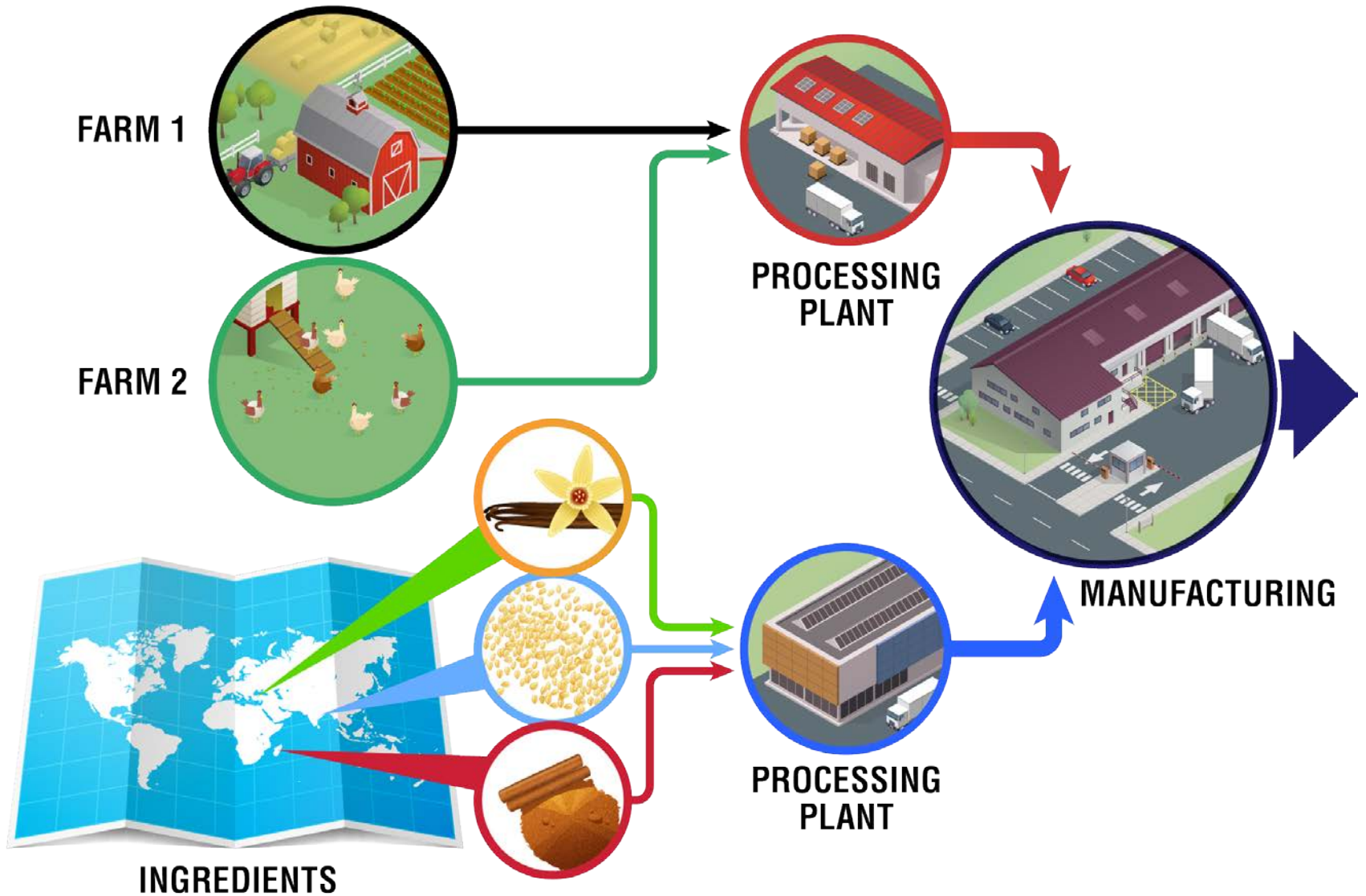


Identifying an Outbreak Vehicle: Lines of Evidence

Three types of evidence used:

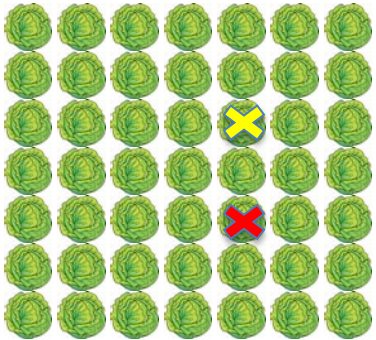
- **Epidemiologic**: association between illness and food exposure
- **Traceback**: suspected food item links back to a common source of contamination
- **Microbiologic/laboratory**: pathogen found in the food, farm or facility



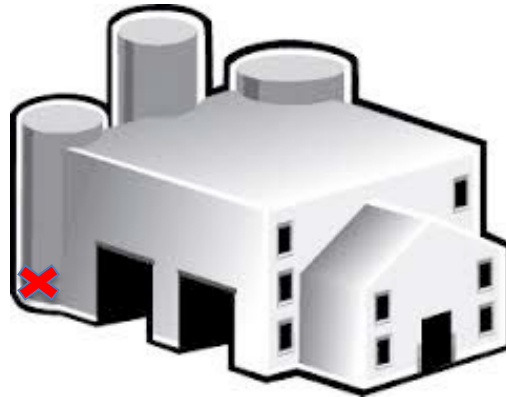
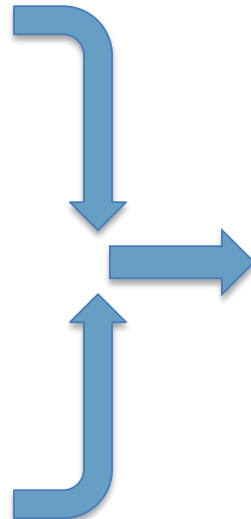
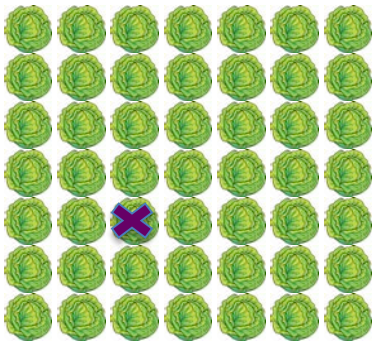


Environmental or Resident Contamination

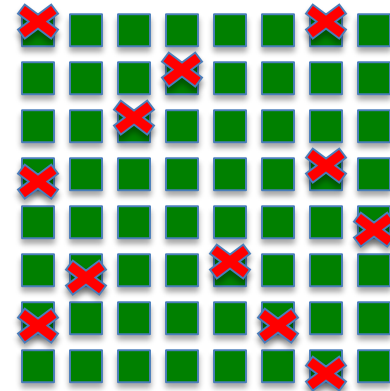
Field 1



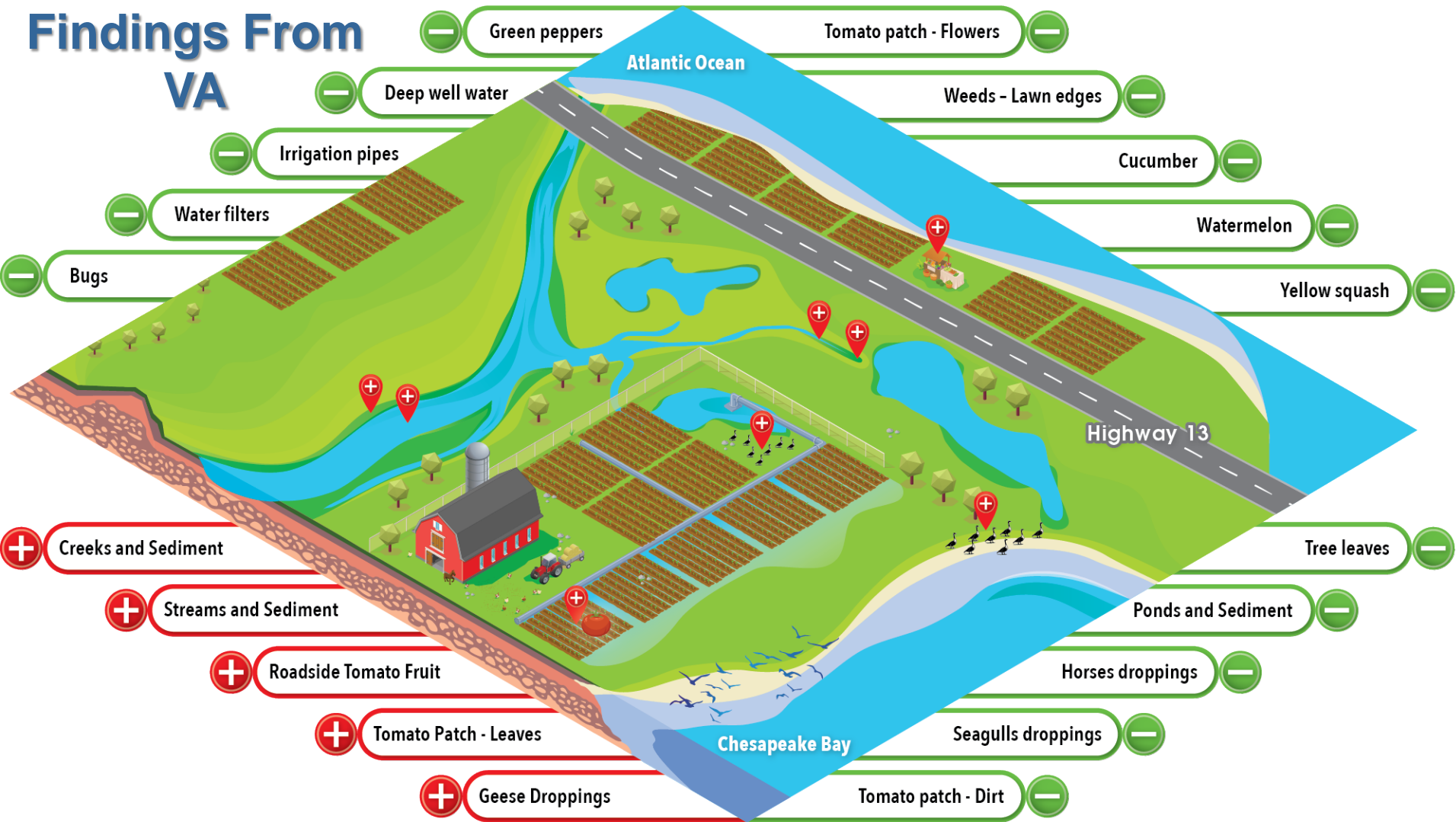
Field 2



Processing facility



Findings From VA



Collected over 500 isolates:

2010-2011: **237**

2012: **137**

2014: **133+**

Key Conclusions from VA surveys Identify Water, Water, and Water



- Sampled Areas:
 - Virginia Tech Agricultural Research and Education Center (AREC): 2009-2011
 - 6 environmental waters and sediments: 2011, 2012, 2014-2015
- *Salmonella* Pattern 61 and other clinically relevant isolates found consistently in waters and sediments at AREC and the 6 other locations
- Environmental waters and sediments are potential reservoirs for *Salmonella*
- Enhanced agricultural practices related to the use of surface waters are important to mitigating *Salmonella* transfer on to crop plants

Bell RL et al. Ecological prevalence, genetic diversity, and epidemiological aspects of Salmonella isolated from tomato agricultural regions of the Virginia Eastern Shore. Front Microbiol. 2015 May 7;6:415.



South American Salmonella Environmentals

Global Water & Food Safety Summit
21 November 2019

Maria Sanchez
James Pettengill
Hugh Rand

•Serotypes Found

45 Infantis

26 Typhimurium

15 IIIb 48:i:z

14 Anatum

12 Enteritidis

10 Paratyphi B

10 Brandenburg

7 I 4[5]12:i:-

4 Rissen

4 Muenchen

4 Mbandaka

4 Manhattan

3 IIIb 58:k:z

2 Johannesburg

2 IIIb 61:i:z

1 -:z10:enz15

1 Senftenberg

1 Predicted Serotype

1 IV 43:z4z23:-

1 Dublin

1 Cerro

1 18:k:z

Cluster Characteristics (N= 28)



	SNPcluster	Water Isolates	Cluster Size
1	PDS000053260.2	2	3
2	PDS000053263.2	2	3
3	PDS000053276.1	4	5
4	PDS000053277.1	2	2
5	PDS000053278.1	2	2
6	PDS000053279.1	3	3
7	PDS000053280.1	2	2
8	PDS000053281.1	4	5
9	PDS000053282.1	4	4
10	PDS000053283.1	3	3
11	PDS000003955.531	10	4330
12	PDS000053285.1	6	4
13	PDS000053286.1	5	5
14	PDS000053288.1	2	2



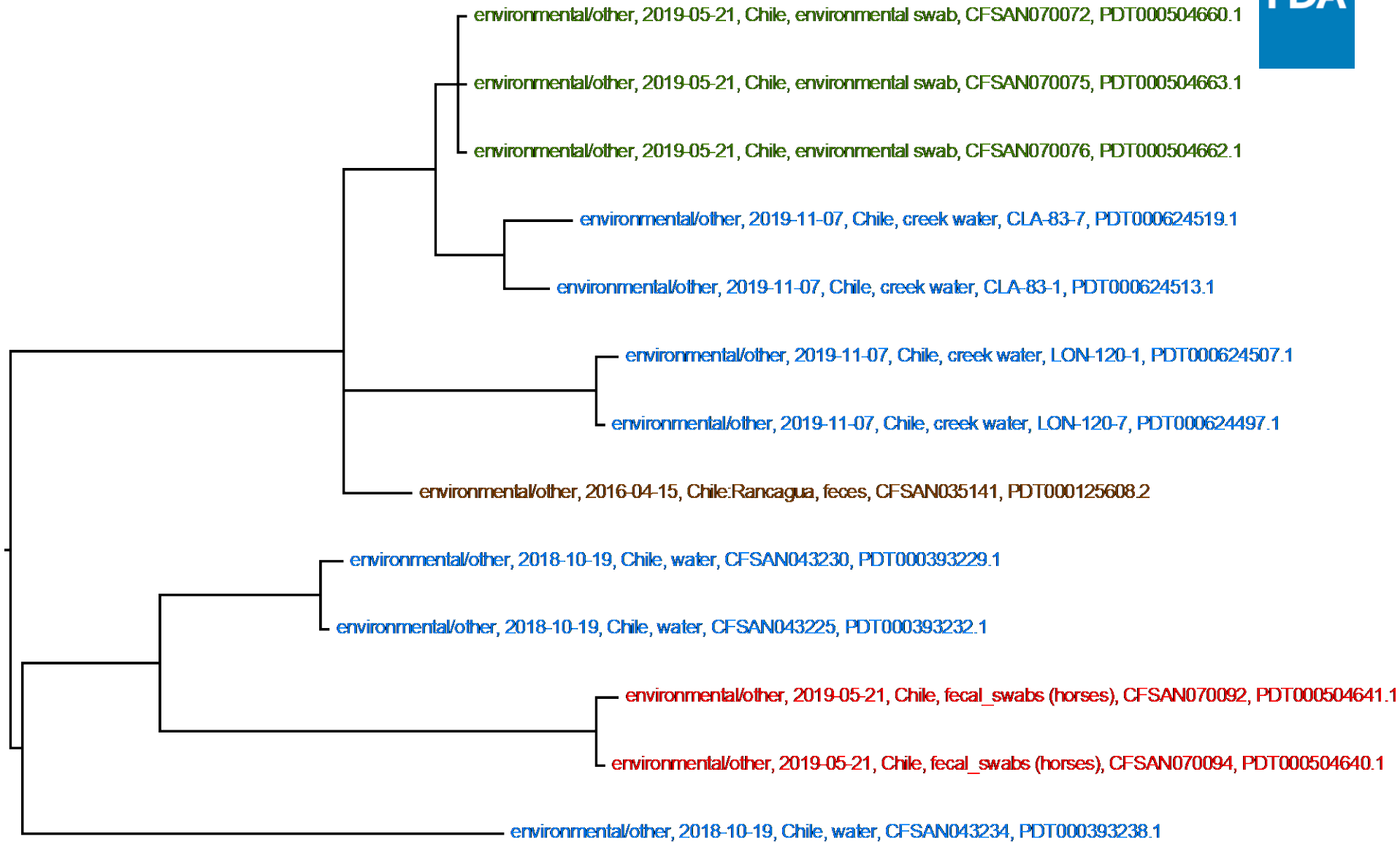
Cluster Characteristics (N= 28)



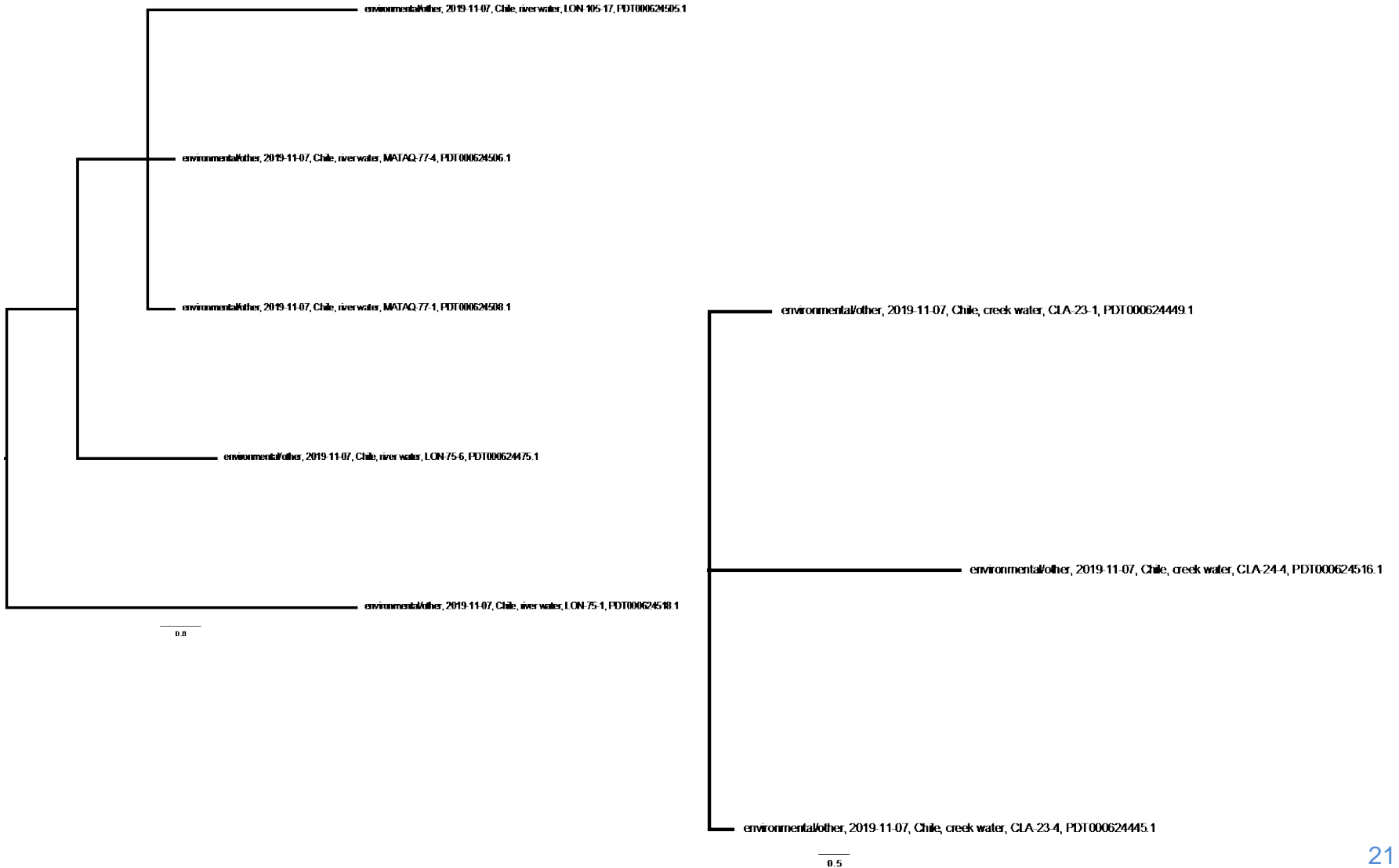
	SNPcluster	Water Isolates	Cluster Size
15	PDS000053289.1	2	2
16	PDS000053290.1	2	2
17	PDS000053291.1	6	7
18	PDS000053292.1	3	3
19	PDS000053293.1	2	2
20	PDS000053294.1	2	2
21	PDS000026867.7	3	55
22	PDS000026991.4	2	7
23	PDS000029248.7	2	24
24	PDS000029912.2	6	8
25	PDS000030058.41	1	89
26	PDS000032393.134	23	861
27	PDS000037691.3	4	13
28	PDS000051343.30	4	2009

SNP Cluster	AMR	BIOCIDES	METAL	VIRULENCE
PDS000003955.533	63	7	56	14
PDS000026867.7	6	0	9	0
PDS000029248.7	4	0	8	0
PDS000029912.2	10	0	15	0
PDS000030058.41	3	0	3	0
PDS000032393.135	32	0	64	0
PDS000037691.3	8	0	12	0
PDS000051343.31	8	0	12	0
PDS000053260.2	3	0	3	0
PDS000053263.2	4	0	6	0
PDS000053277.1	2	0	3	0
PDS000053278.1	0	0	2	4
PDS000053279.1	6	0	9	0
PDS000053280.1	6	0	6	0
PDS000053281.1	19	0	9	0
PDS000053282.1	4	0	6	0
PDS000053283.1	6	0	9	0
PDS000053286.1	10	0	15	0
PDS000053288.1	4	0	6	0
PDS000053289.1	0	0	2	4
PDS000053290.1	4	0	6	0
PDS000053291.1	12	0	18	0
PDS000053292.1	4	0	6	0
PDS000053293.1	4	0	6	0
PDS000053294.1	4	0	6	0

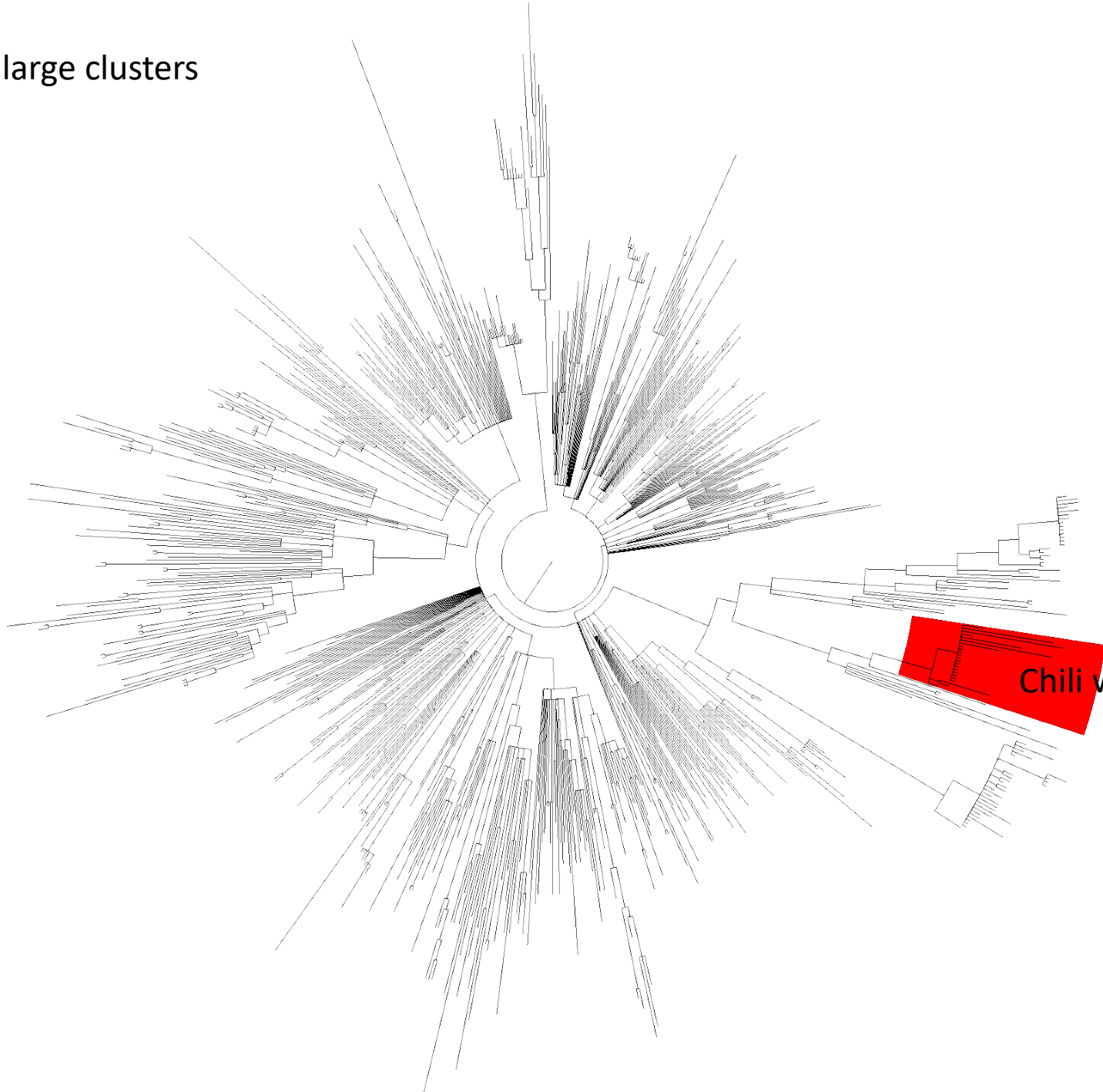
Diversity of isolates from Chile



Many small clusters



But also large clusters



Chili water isolates

www.ncbi.nlm.nih.gov/pathogens/



Pathogen Detection BETA

NCBI Pathogen Detection integrates bacterial pathogen genomic sequences originating in food, environmental sources, and patients. It quickly clusters and identifies related sequences to uncover potential food contamination sources, helping public health scientists investigate foodborne disease outbreaks.

[Find isolates now!](#)

Explore the Data

Species	New Isolates	Total Isolates
Salmonella enterica	124	53,003
E.coli and Shigella	1	19,861
Listeria monocytogenes	20	12,267
Campylobacter jejuni	0	4,309
Acinetobacter baumannii	0	2,651
Klebsiella pneumoniae	1	2,212
Enterobacter	0	1,151
Vibrio parahaemolyticus	0	817

Learn More

- [About](#)
- [FAQ](#)
- [Antimicrobial Resistance](#)
- [Contributors](#)

Data Resources

- [Isolates Browser](#)
- [Antimicrobial resistance reference gene database](#)
- [Isolates with antibiotic resistant phenotypes](#)
- [Beta-lactamase resources](#)
- [Download analysis results \(FTP\)](#)

Submit

- [How to submit data](#)
- [How to submit antibiotic resistance phenotypes](#)
- [How to submit beta-lactamases](#)
- [NCBI Submission Portal](#)

Publicly released metadata

Food/environmental Submission

Pathogen: environmental/food/other sample from

Listeria monocytogenes

BioSample: SAMN02709234;

Sample name: FDA00007620

Organism Listeria monocytogenes

cellular organisms; Bacteria; Firmicutes; Bacilli;
Bacillales; Listeriaceae; Listeria

Pathogen: environmental/food/other; v.1.0

strain FDA00007620

host disease missing

latitude and longitude missing

collection date 3/26/2012

isolate CFSAN003790

geographic location Italy

isolation source moliterno al tartufo
cheese

collected by FDA

PFGE_SecondaryEnzyme_pattern GX6A12.0280

PFGE_PrimaryEnzyme_pattern GX6A16.0085

Submission

FDA, Justin Payne; 2014-03-27

ID: 2709234

Clinical Sample Submission

**Pathogen: clinical or host-associated sample
from Salmonella enterica**

BioSample: SAMN02927343;

Sample name:/2013K-0563

Organism Salmonella enterica

cellular organisms; Bacteria; Proteobacteria;
Gammaproteobacteria; Enterobacteriales;

Enterobacteriaceae; Salmonella

Pathogen: clinical or host-associated; version 1.0

strain 2013K-0563

collected by CDC

collection date Missing

geographic location USA

host Missing

host disease Missing

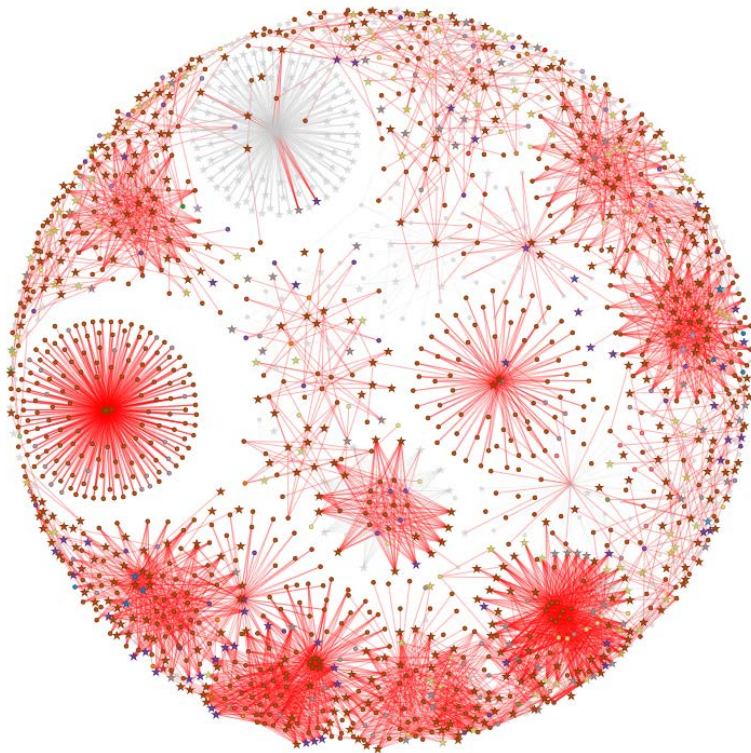
isolation source Missing

latitude and longitude Missing

Submission

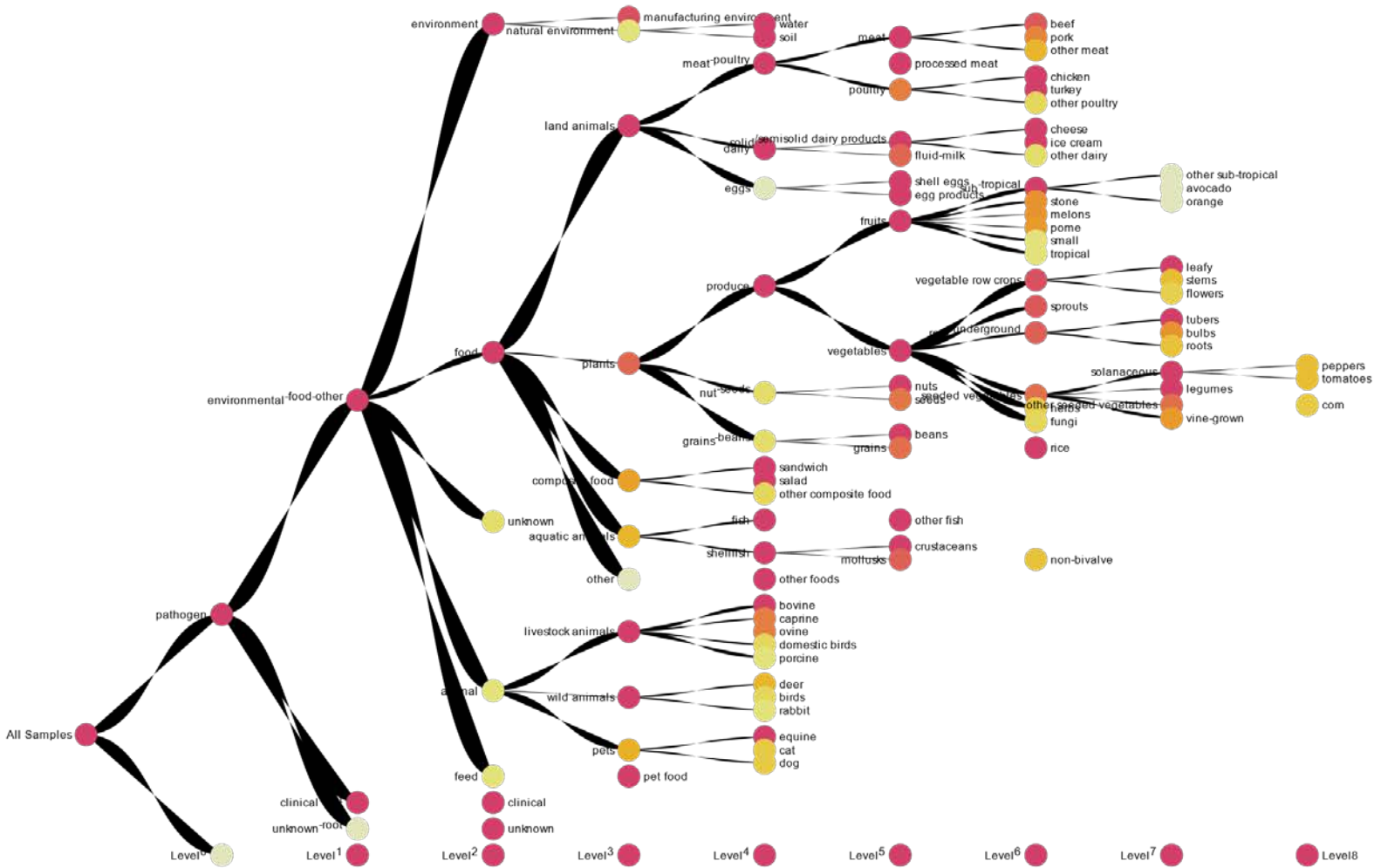
Pulsenet, Eija Trees; 2014-07-18

ID: 2927343



*Leveraging GenomeTrakr &
NCBI Pathogen Detection WGS
Data to Enhance*

- Risk Assessment
- Attribution
- Large-scale Epidemiology studies



GenomeGraphR: WGS data integration, analysis, and visualization for risk assessment and management: <https://fda-riskmodels.foodrisk.org/genomegraphr/>
 Moez Sanaa, Régis Pouillot, Francisco J Garces-Vega, Errol Strain, Jane M Van Doren doi: <https://doi.org/10.1101/495309> 2018.

Size of Dots

1 2 10

1 2 3 4 5 6 7 8 9 10

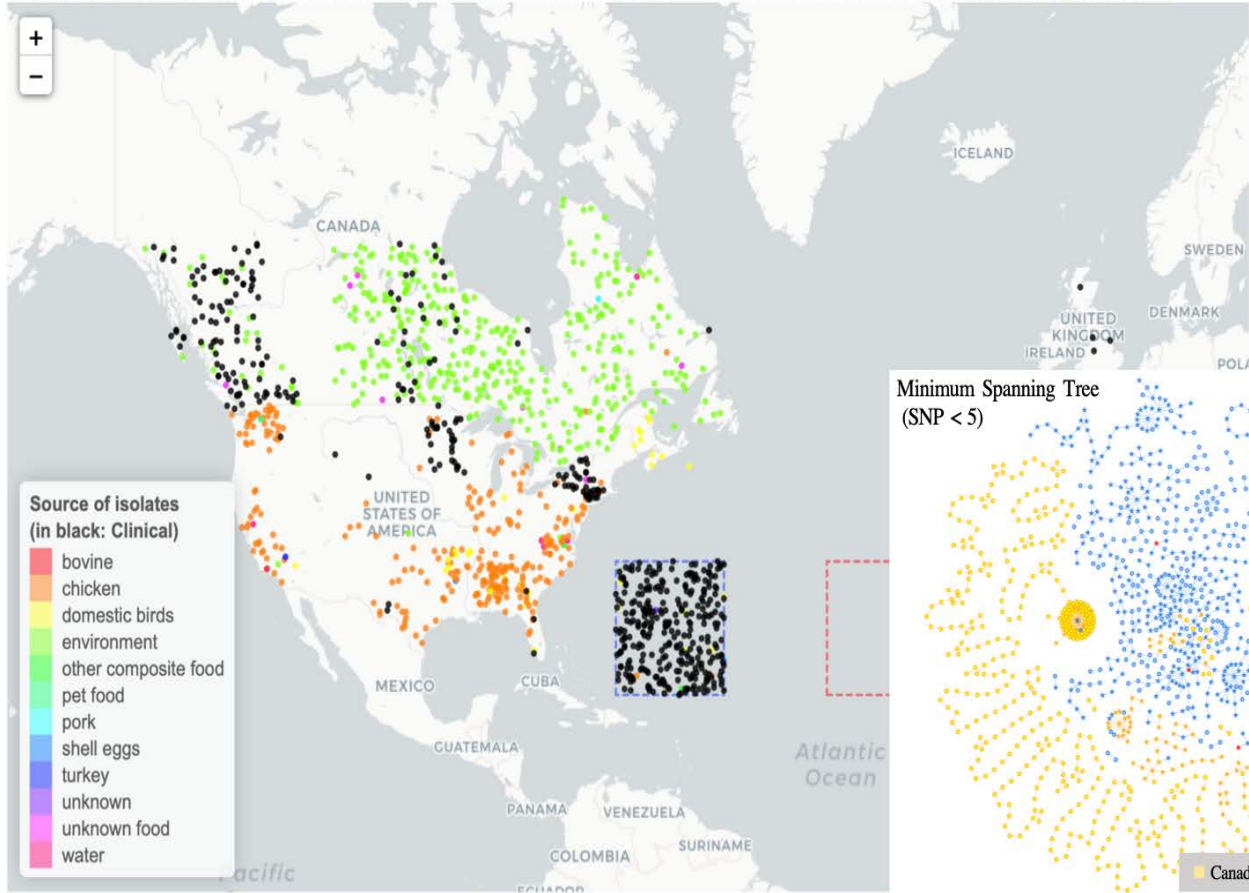
Cumulate data over years

Year of Collection/Creation

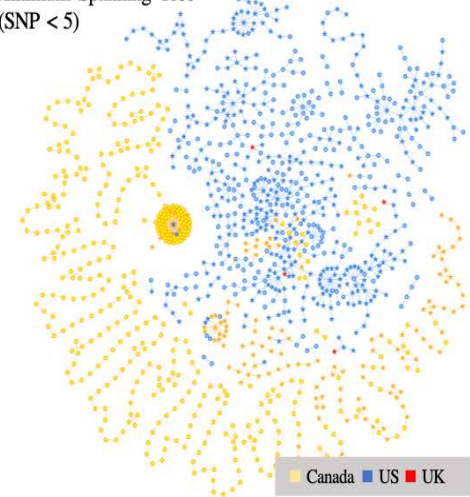
2001 2018

2001 2005 2009 2013 2017

Warnings: the dots are placed at random within the limits of the state (United States) or the country. The position of each dot doesn't represent the actual location of sampling. Strains from the US not assigned to a specific State are placed in the blue square. Strains not assigned to a specific country are placed in the red square.

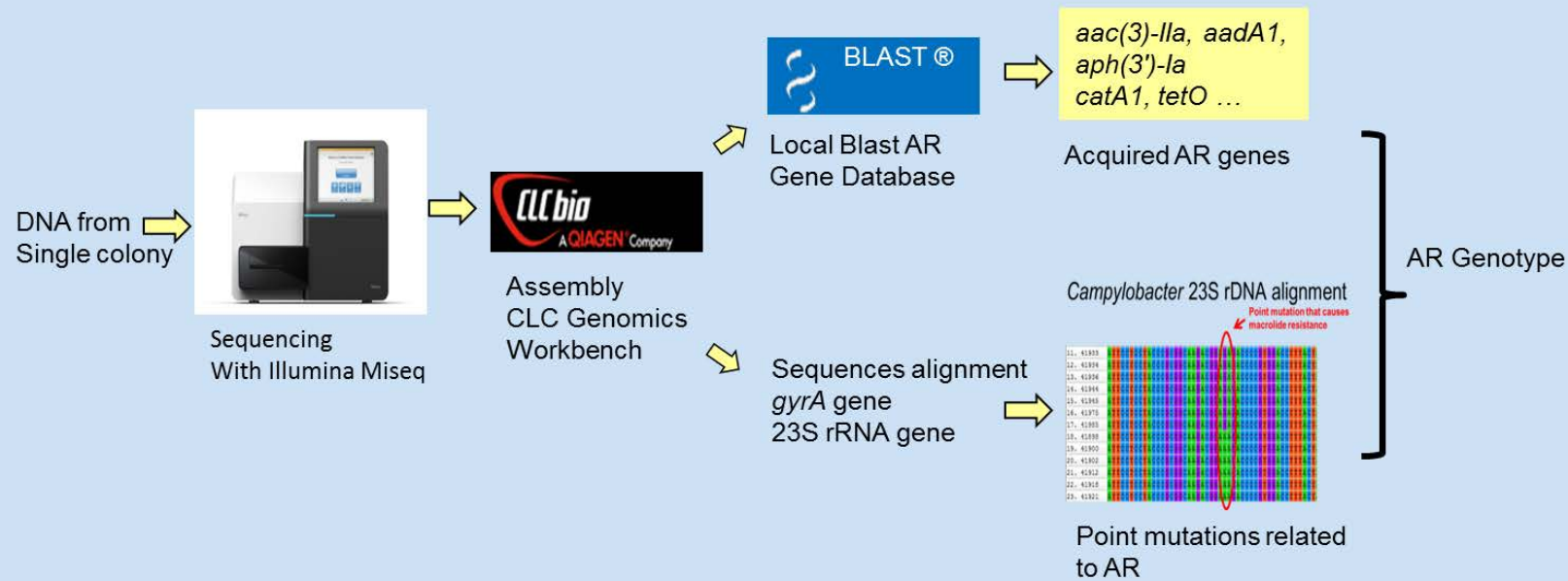


Minimum Spanning Tree (SNP < 5)





From WGS to Antibiotic Resistance Genotype



RESISTOME TRACKER

Salmonella

Select an icon or alert below to get started.



CUSTOMIZE



COMPARE



DISCOVER



EXPLORE

ALERTS

There are **11** records with **2** flagged genes uploaded in the last 30 days



Manuscripts using structured metadata ontologies, machine learning and artificial intelligence.

Nguyen M, Long SW, McDermott PF, Olsen RJ, Olson R, Stevens RL, Tyson GH, Zhao S, Davis JJ. 2019. **Using machine learning to predict antimicrobial MICs and associated genomic features for nontyphoidal *Salmonella***. J Clin Microbiol 57:e01260-18. <https://doi.org/10.1128/JCM.01260-18>.

Zhang et al. 2019. **Zoonotic Source Attribution of *Salmonella enterica* Serotype Typhimurium Using Genomic Surveillance Data, United State**. Emerg Infect Dis. 25(1): 82–91. doi: 10.3201/eid2501.180835

Adaptations of particular interest to food safety specialists:

- | | |
|-----------------------------------|---|
| (1) Thermal tolerance | (18) Surface water fitness |
| (2) Dessication resistance | (19) In vivo plant migratory fitness |
| (3) Osmotic/Ionic tolerance | (20) Soil fitness |
| (4) Quat resistance | (21) Capsaicin resistance |
| (5) Chlorine resistance | (22) Swarming |
| (6) Biofilm persistence | (23) Trans-ovarian poultry colonization |
| (7) Surface adherence | (24) Fecal persistence (poultry) |
| (8) Antibiotic resistance | (25) Yolk content invasion |
| (9) Antimicrobial resistance | (26) Multidrug resistance |
| (10) Ecological fitness | (27) External amoeba harborage |
| (11) Heavy metal resistance | (28) Internal amoeba harborage |
| (12) Metabolic persistence | (29) Acyl-homoserine lactone (AHL) |
| (13) Enhanced hydrophobic fitness | (30) KatE stationary-phase catalase |
| (14) Produce invasiveness | (31) In vivo migratory fitness |
| (15) Flower invasiveness | (32) RDAR phenotype |
| (16) Root system invasiveness | (33) The 'Weltevreden' type |
| (17) Acid resistance | (34) Persistence within the tomato** |

A partial gene list is now incorporated into AMRFinderPlus v3.2 (<https://github.com/ncbi/amr/wiki>) and the Reference Gene Catalog as of 2019-10-31 (<https://www.ncbi.nlm.nih.gov/pathogens/isolates#/refgene/>)

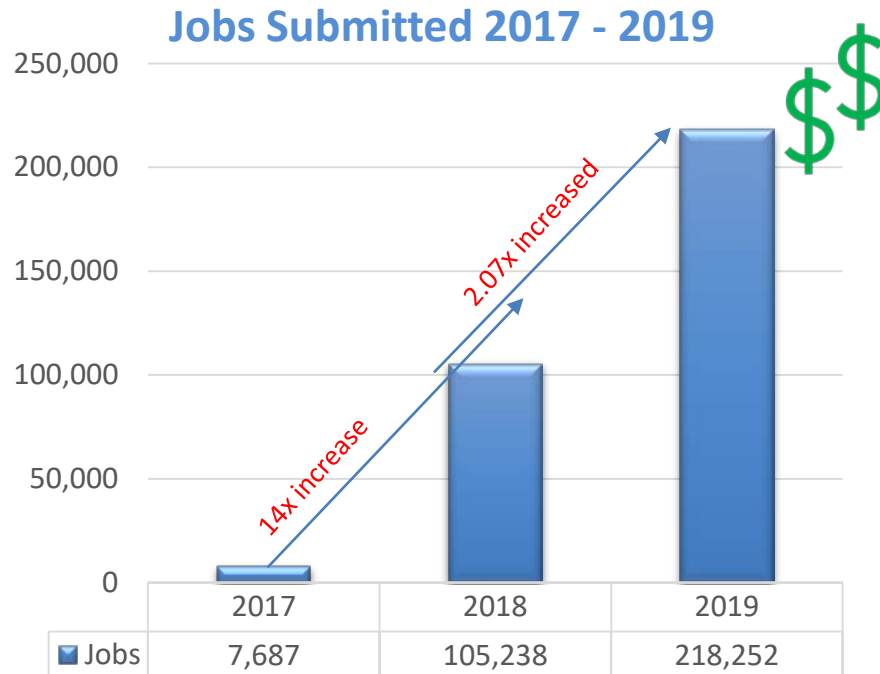


- **2019 Objective: Increase Jobs Submitted to Galaxy by 1.5 times to 160,000**



- **End of Year Status: Achieved!**

- ✓ From 2017 to 2018, number of jobs submitted increased by 14x
- ✓ From 2018 to 2019, number of jobs submitted increased by 2.07x





- **2019 Objective: Increase Jobs Submitted to Galaxy by 1.5 times to 160,000**
-



- **Benefit / Impact:**

- ✓ **GalaxyTrakr Training Environment**
 - Deployed a separate training environment to support various GalaxyTrakr training events without impacting production
- ✓ **Updated to AWS S3**
 - Updated object storage services (from AWS EFS to AWS S3) allowing an increase in user storage quotas AND additional 90% reduction in storage costs (\$.30 to \$.003 per GB)
- ✓ **Increased Number of Users:**
 - Current Number of Registered Users: 522 (10% increase since last quarterly meeting: 475 registered users)
- ✓ **Over 70 tools and 11 workflows available on GalaxyTrakr**
 - Increase in tools supports overall mission objectives and expands the overall GalaxyTrakr community

- Future Projects
- GenomeTrakr Latin America



For more discussions please seek out Gonzalez-Escalona, Narjol <Narjol.Gonzalez-Escalona@fda.hhs.gov>; Stevens, Eric <Eric.Stevens@fda.hhs.gov>; Cornell, Jason <Jason.Cornell@fda.hhs.gov>

Acknowledgements

- **FDA**
 - Center for Food Safety and Applied Nutrition
 - Center for Veterinary Medicine
 - Office of Regulatory Affairs
- **National Institutes of Health**
 - National Center for Biotechnology Information
- **State Health and University Labs**
 - Alaska
 - Arizona
 - California
 - Florida
 - Hawaii
 - Maryland
 - Minnesota
 - New Mexico
 - New York
 - South Dakota
 - Texas
 - Virginia
 - Washington
- **USDA/FSIS and ARS**
- **CDC**
 - Enteric Diseases Laboratory
- **INEI-ANLIS “Carlos Malbran Institute,” Argentina**
- **Centre for Food Safety, University College Dublin, Ireland and Irish FSA**
- **Melbourne (FSA). Australia**
- **Public Health England, UK**
- **Institute for Food Safety and Health (IFSH)**
- **WHO and FAO**
- **Illumina**
- **Pac Bio**
- **Other independent collaborators**



