

REFINING SAMPLING AND ANALYSIS APPROACHES TO ADVANCE UNDERSTANDING OF THE MICROBIOLOGICAL RISKS OF AGRICULTURAL WATER REUSE



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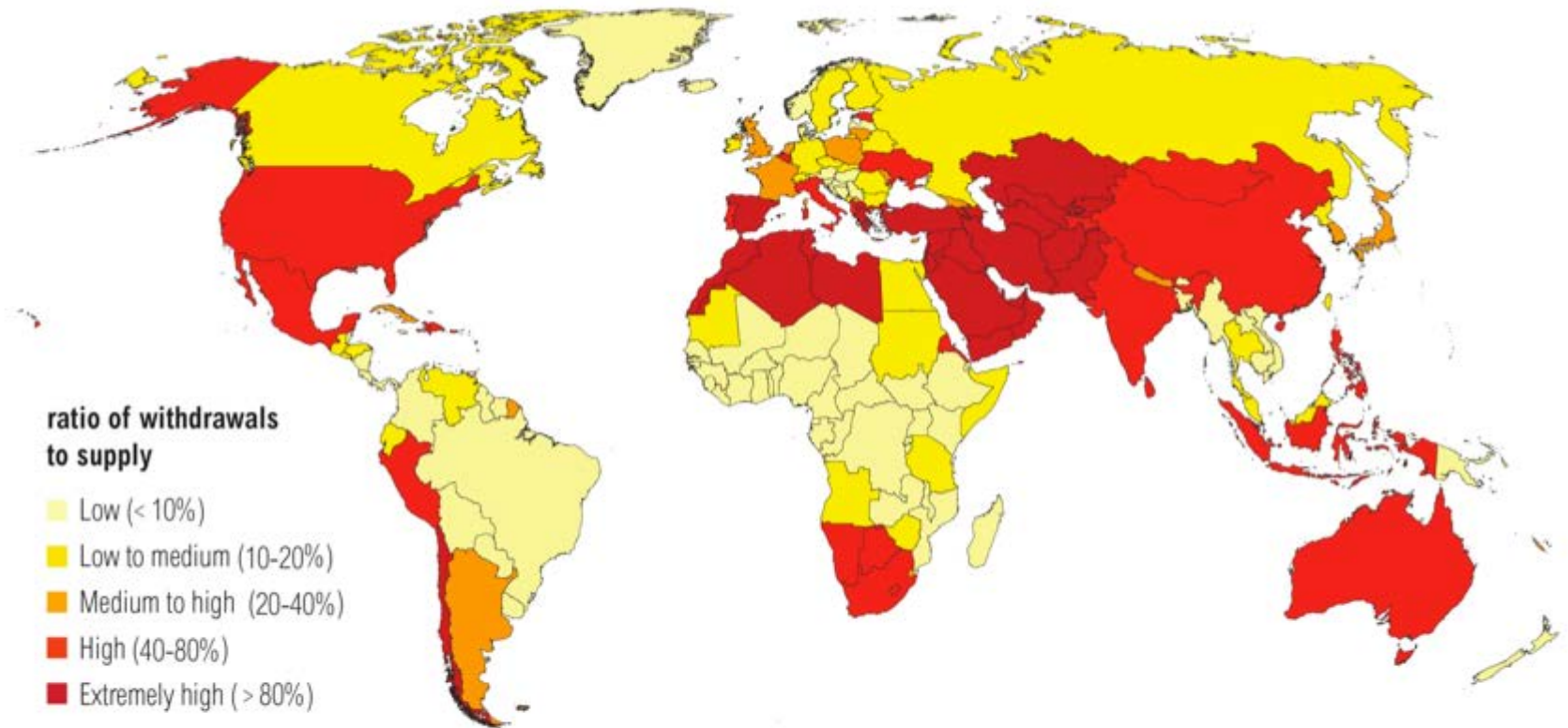
PRESENTATION OUTLINE

- Global water scarcity and use of recycled water on food crops
- Mission and activities of CONSERVE Center of Excellence
- Refining sampling and analysis approaches for recycled water
 - Comparing sampling frequency and sample volumes
 - Employing whole genome sequencing
 - Coupling DNA-labeling and sequencing approaches



INCREASING WATER SHORTAGES THREATEN IRRIGATION WATER RESOURCES

Water Stress by Country: 2040



NOTE: Projections are based on a business-as-usual scenario using SSP2 and RCP8.5.

For more: ow.ly/RiWop

RECYCLED WATER: AN IMPORTANT PART OF THE SOLUTION

- Recycled water = advanced treated wastewater or greywater = reclaimed water = water reuse
- “One Water Approach” = All water has value and should be managed in a sustainable, inclusive, integrated way
- “There is no bad water”



Photo: ClimateTechWiki



Source: Carollo.com



Orchards being irrigated with untreated wastewater in Riverside, CA, 1890-1900.

California Historical Society Collection, 1860-1960

AGRICULTURAL IRRIGATION ACCOUNTS FOR ~30% OF RECYCLED WATER USE IN THE U.S.

Table 3-3. Nationwide reuse summaries of reclaimed water use in agricultural irrigation (adapted from Bryk et al., 2011)

State	Annual Agricultural Reuse Volume	
	mgd	1000 ac-ft/yr
Arizona	23	26
California	270	303
Colorado	2.97	3
Florida	256	287
Idaho	0.27	0.3
North Carolina	1.0	1
Nevada	13.4	15
Texas	19.4	22
Utah	0.81	1
Washington	0.02	0.03
Wyoming	0.89	1

- Number of US States with rules, regulations or guidelines addressing water reuse on:
 - Food crops: 27 states
 - Processed food and non-food crops: 43 states
- Rules, regulations and guidelines dictate **water quality requirements**

FOOD SAFETY MODERNIZATION ACT (FSMA)

- FSMA Produce Safety Rule
 - Strict water quality criteria for irrigation water “that is directly applied to growing produce...”
 - *E. coli* standard:
 - Baseline microbial water quality profile for untreated surface water: \geq 20 samples over 2-4 years
 - Geometric mean (GM) < 126 CFUs of generic *E. coli* per 100 mL
 - Statistical threshold value (STV) < 410 CFUs of generic *E. coli* per 100 mL

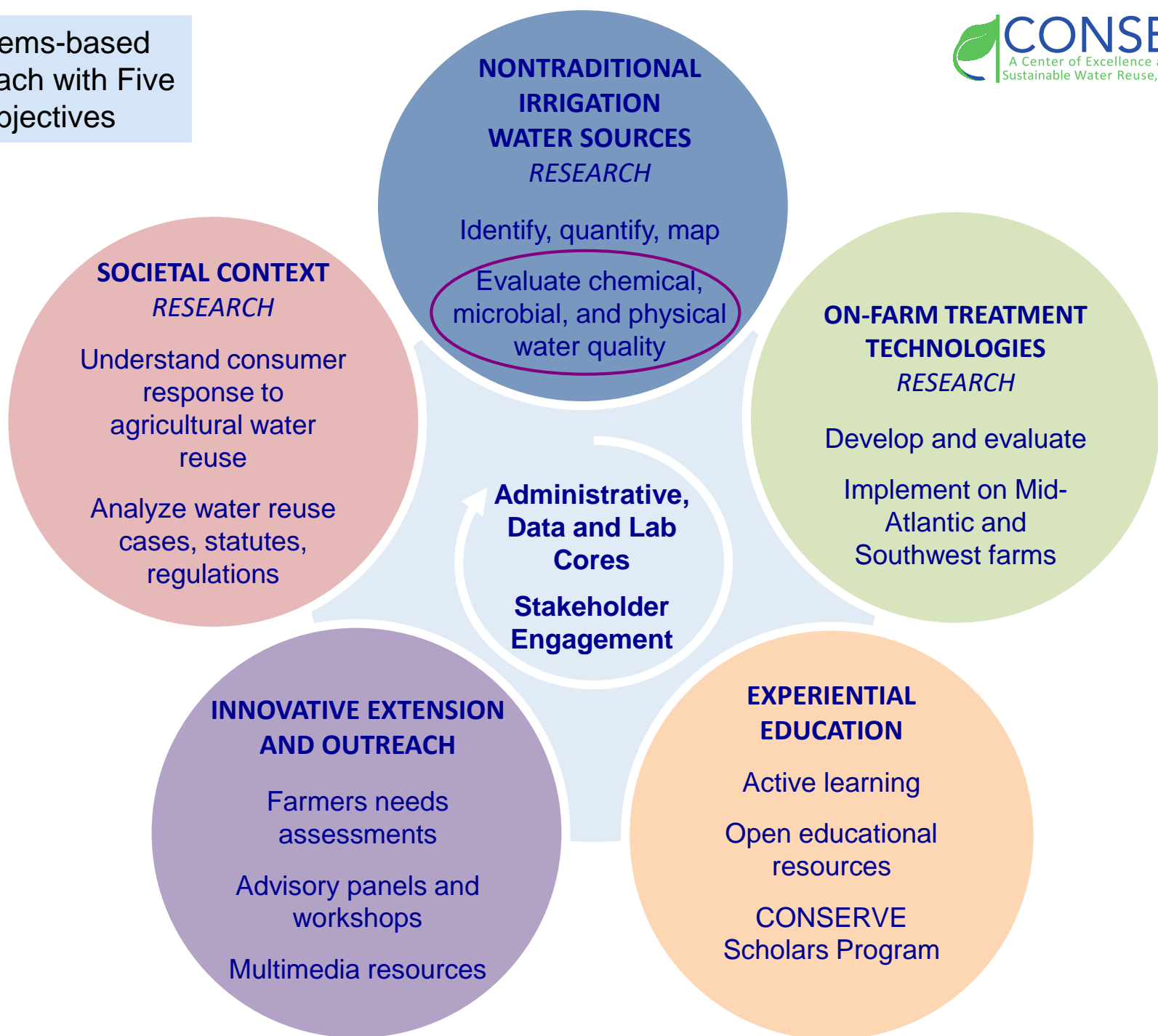


Sustainable on-farm water treatment solutions are needed to enable growers to use recycled (or nontraditional) water sources for food crop irrigation.

Our Mission: To facilitate the adoption of transformative on-farm water treatment solutions that enable the safe use of nontraditional irrigation water on food crops.



Systems-based
Approach with Five
Objectives



Nontraditional Irrigation Water Quality

(Co-PIs: Sapkota, Kniel, Sharma, Micallef, Hashem, Gerba, Ravishankar, Rock, Parveen, May, Sapkota, Mongodin, Colwell, Pop)

2 Year, Bi-weekly Field Sampling Effort Completed



22 field sites sampled in the Mid-Atlantic and Southwest

>5,000 water samples collected, processed, analyzed for bacterial indicators, pathogens, antibiotic-resistant bacteria, pharmaceuticals, and sent for 16S rRNA sequencing and metagenomic shotgun sequencing

>500 *Salmonella* isolates Whole Genome Sequenced by FDA GenomeTrakr Program



E. coli Findings: Mid-Atlantic Sites October 2016-October 2018



SARAH ALLARD

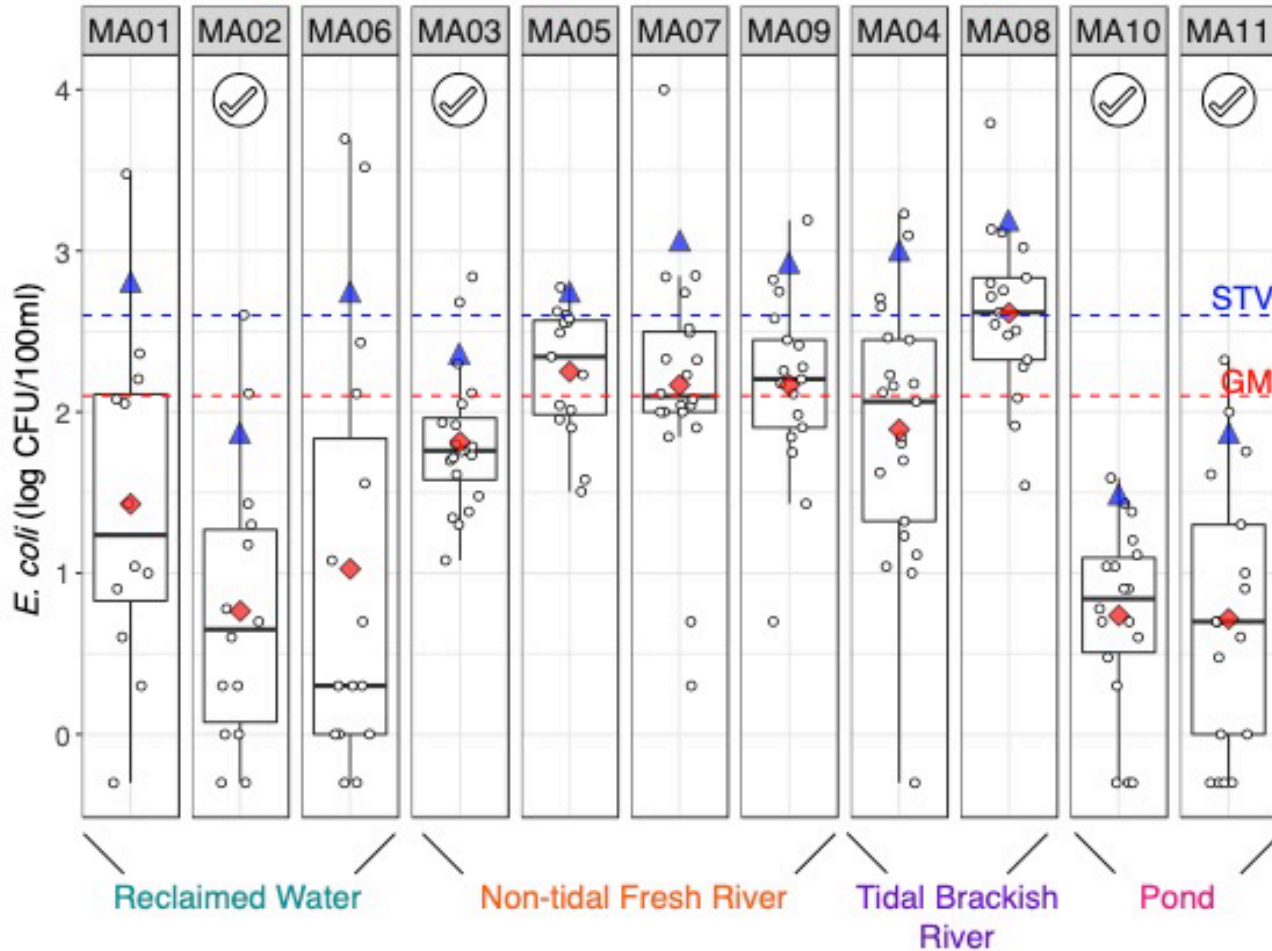
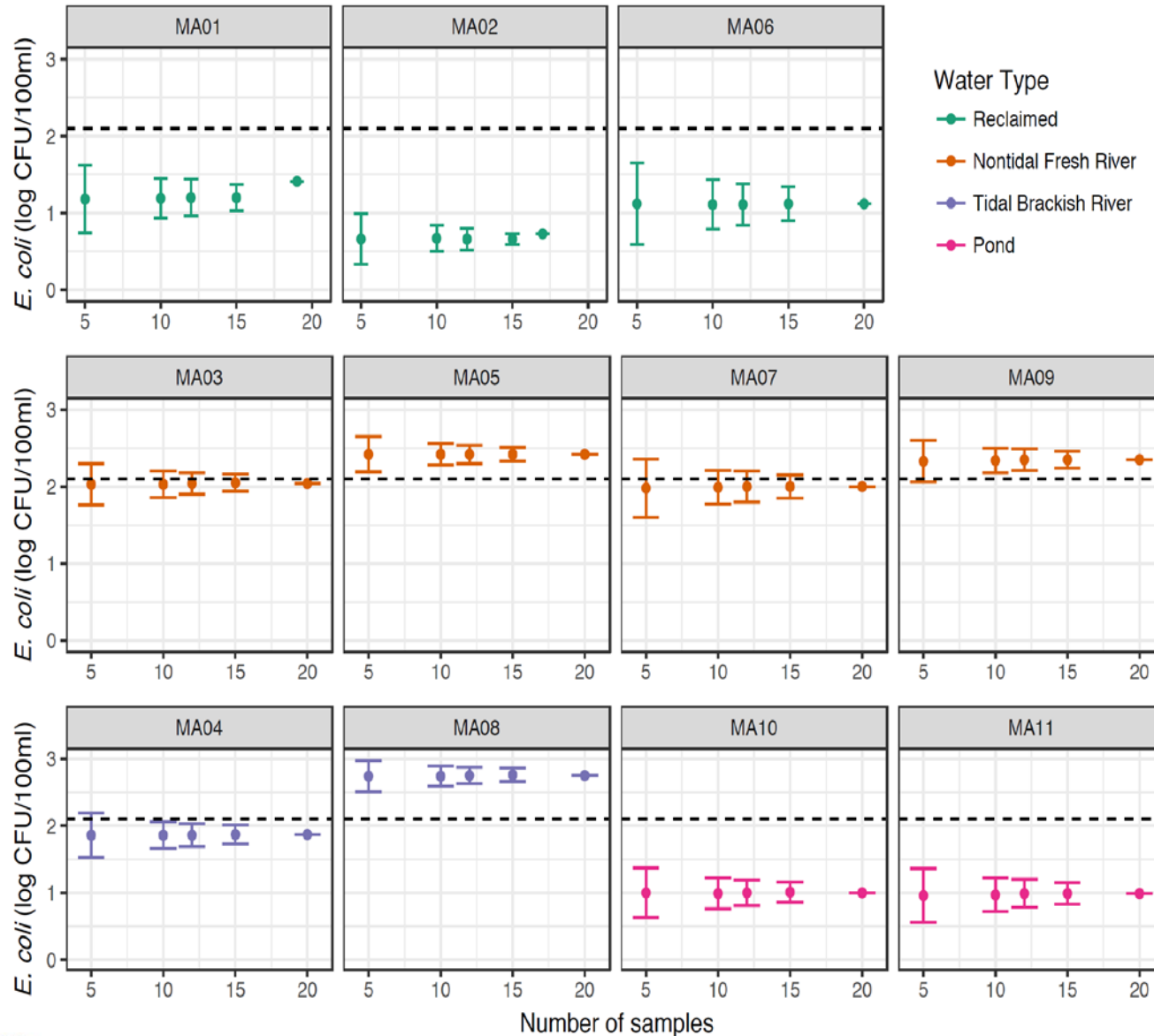


Fig 1: *E. coli* geometric mean (GM, red diamond) and statistical threshold values (STVs, blue triangle) in irrigation water samples collected during the growing season (on sampling days with <0.1 cm precipitation in the previous 24 h) in comparison to standards of the Food Safety Modernization Act, Produce Safety Rule.

Impact of Sampling Frequency on *E. coli* Geometric Means



Impact of Sample Volume on Pathogen Detection

Table 3: Number (percentage) of total sampling events at each site where each water volume filtered contained *Salmonella* spp. or *L. monocytogenes*.

Site	Water type	# Sampling event	<i>Salmonella</i> spp.			<i>L. monocytogenes</i>		
			0.1 L	1 L	10 L	0.1 L	1 L	10 L
MA04	River	34	17 (50%)	16 (47.1%)	27 (79.4%)	9 (26.5%)	6 (17.6%)	14 (41.2%)
MA05	River	32	8 (25%)	15 (46.9%)	25 (78.1%)	25 (78.1%)	29 (90.6%)	29 (90.6%)
MA06	Reclaimed	25	2 (8%)	5 (20%)	8 (32%)	2 (8%)	2 (8%)	2 (8%)
MA10	Pond	35	1 (2.9%)	2 (5.7%)	7 (20%)	2 (5.7%)	2 (5.7%)	3 (8.6%)
MA11	Pond	34	2 (5.9%)	4 (11.8%)	10 (29.4%)	1 (2.9%)	2 (5.9%)	3 (8.8%)
MA12	Produce wash	10	5 (50%)	4 (40%)	6 (60%)	1(10%)	1 (10%)	1 (10%)

Recovering 10L Water Samples Significantly Improved the Likelihood of Detection for Both *Salmonella* spp. and *Listeria monocytogenes*

Pathogen	Volume Comparison	Increase in Likelihood of Recovery	<i>p</i> -value
<i>Listeria monocytogenes</i>	1L vs 0.1L	1.2	0.894
	10L vs 0.1L	4.8	0.012
	10L vs 1L	3.9	0.037
<i>Salmonella</i> spp.	1L vs 0.1L	1.7	0.194
	10L vs 0.1L	43.5	<0.0001
	10L vs 1L	25.5	<0.0001

Whole Genome Sequencing of *Salmonella* Isolates Revealed the Presence of 21 Serovars

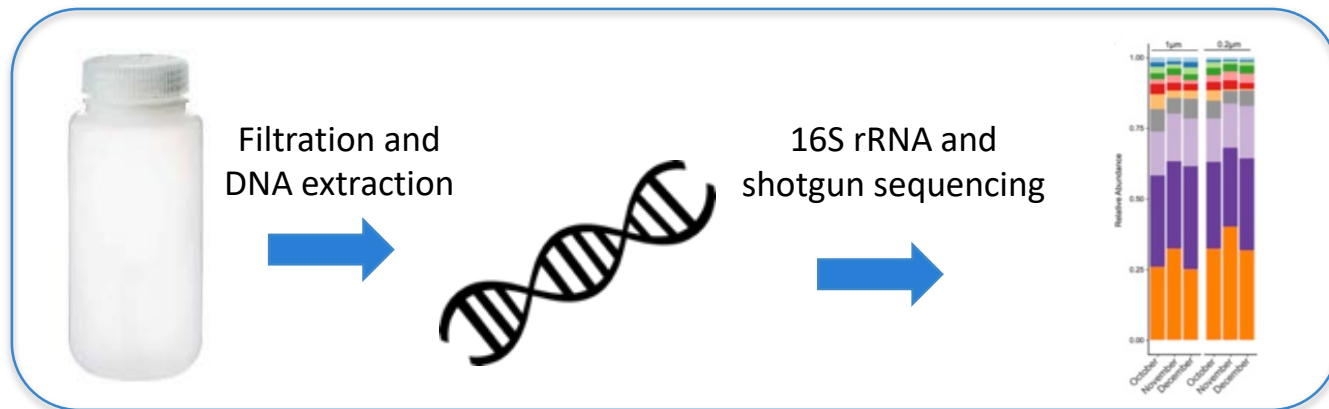
<u>MA 04 (River)</u>	<u>MA 05 (River)</u>	<u>MA 06 (Recycled)</u>	<u>MA 10 (Pond)</u>	<u>MA 11 (Pond)</u>	<u>MA 12 (Processing)</u>
Newport Bareilly Javiana Typhimurium 4:i:- Infantis Thompson Anatum Enteritidis IV44:z36,[z38]:- Johannesbur	Newport Bareilly Javiana 4:i:- Norwich Bovismorbificans Give Oyonnax Illa 48:g,z51:-	Newport Montevideo	Bareilly	Newport Bareilly Typhimurium Thompson Berta	Newport Javiana Typhimurium Infantis Norwich Liverpool Mbandaka

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Fig 1: *Salmonella* serovars isolated from recycled and untreated surface waters and whole genome sequenced through the FDA GenomeTrakr Network.

“Who” else is there and what are they doing?

- Important to culture fecal indicator bacteria (e.g. *E. coli*) and pathogens from irrigation water sources
- Bacterial, culture-based work provides only one part of the overall picture of microbial water quality
- DNA-based approaches can improve our understanding of total microbial communities (taxonomy and function) present in water



Bacterial Diversity and Antibiotic Resistance Genes In Recycled and Untreated Surface Waters



JESSICA CHOPYK

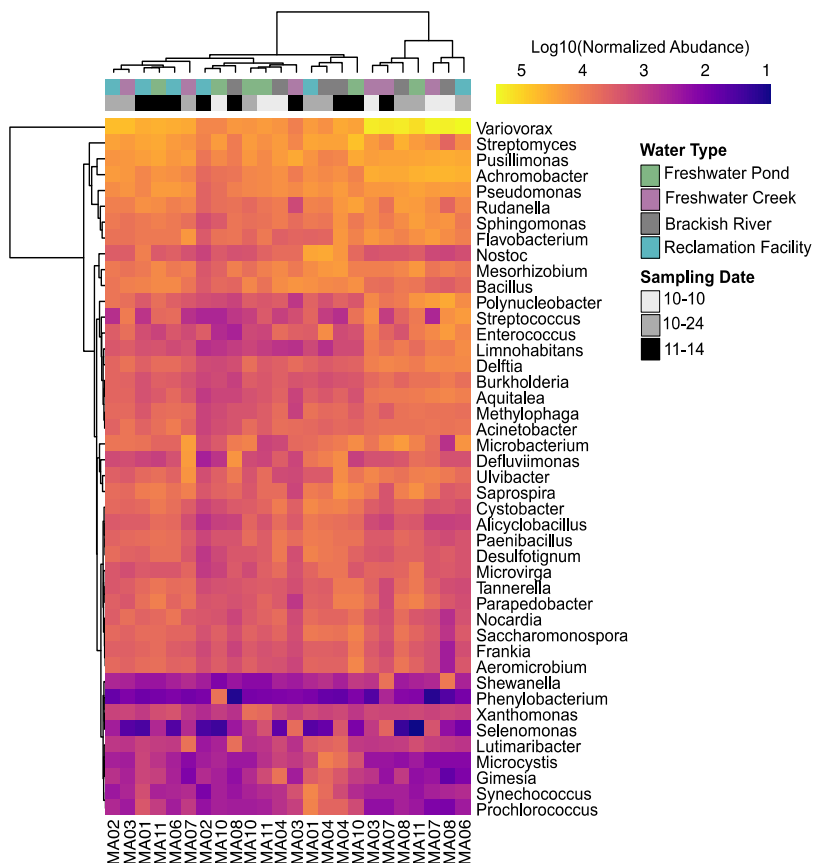


Figure 2: Taxonomic heatmap of the bacterial communities present in recycled and untreated surface water sites by sampling date. Heatmap based on the log-transformed normalized abundance of the most dominant genera (>1% in at least one sample).

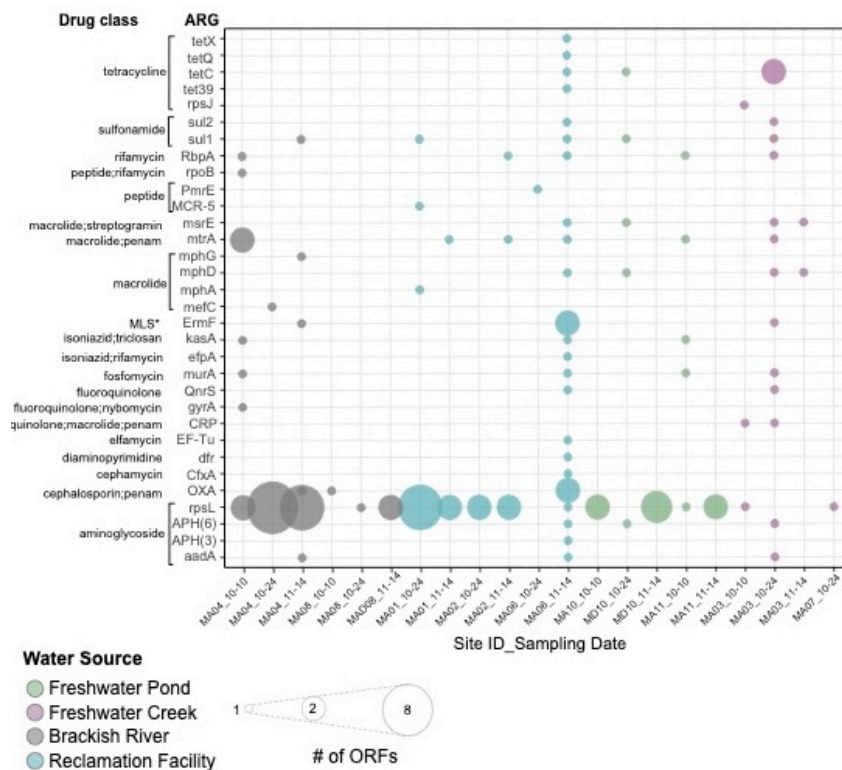


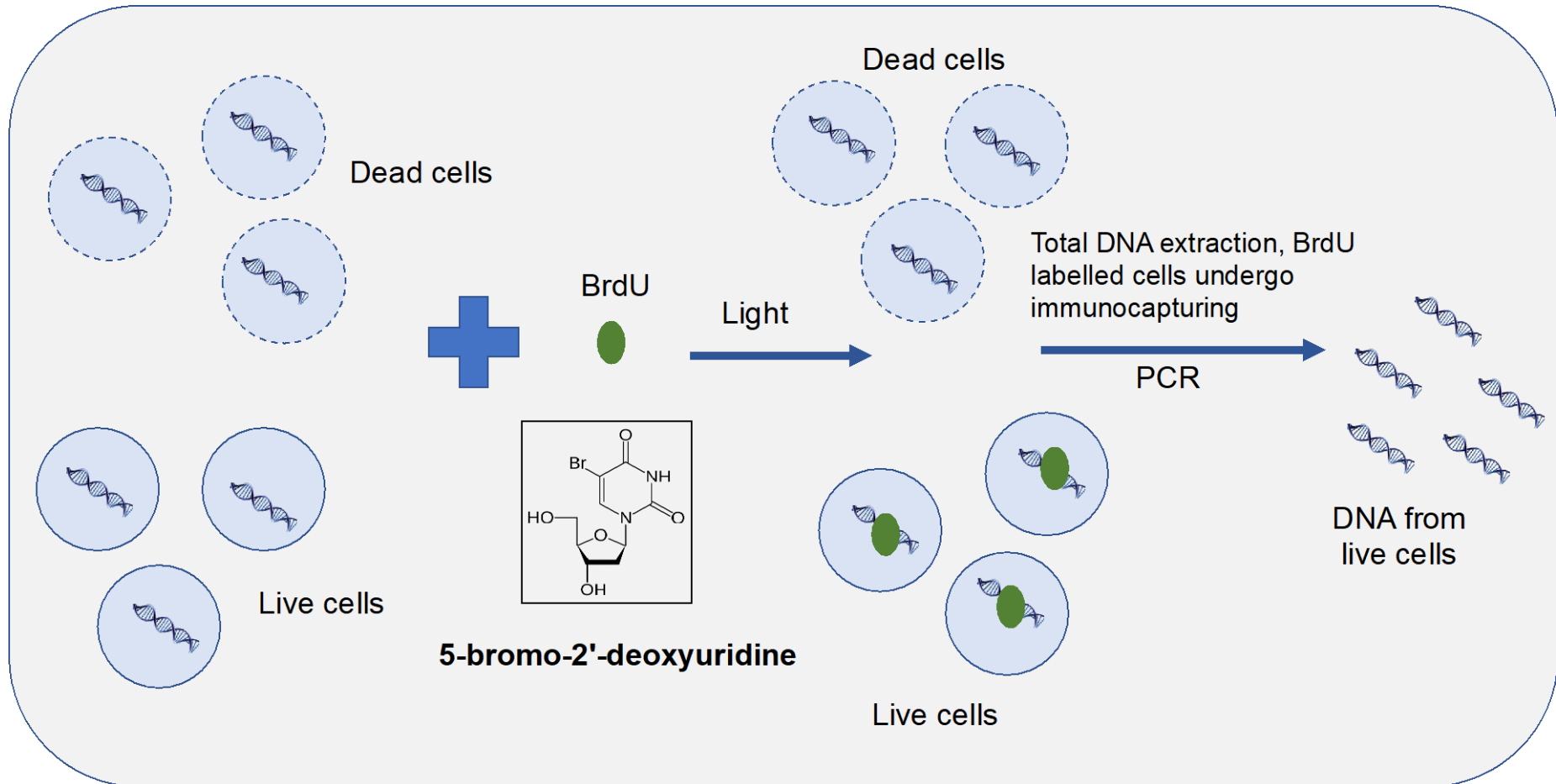
Figure 4: Antibiotic resistance genes (ARGs) predicted in recycled and untreated surface water sites by sampling date. Dotplot showing the ARG-like ORFs present at each water site, with the size of each dot equivalent to the number of translated ORFs with homology to each ARG listed on the y-axis, and the color representative of the water type.

Are they alive (metabolically-active)?



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Coupling DNA-Labeling and Sequencing Approaches



Identifying Metabolically-active Bacterial Communities in Irrigation Water



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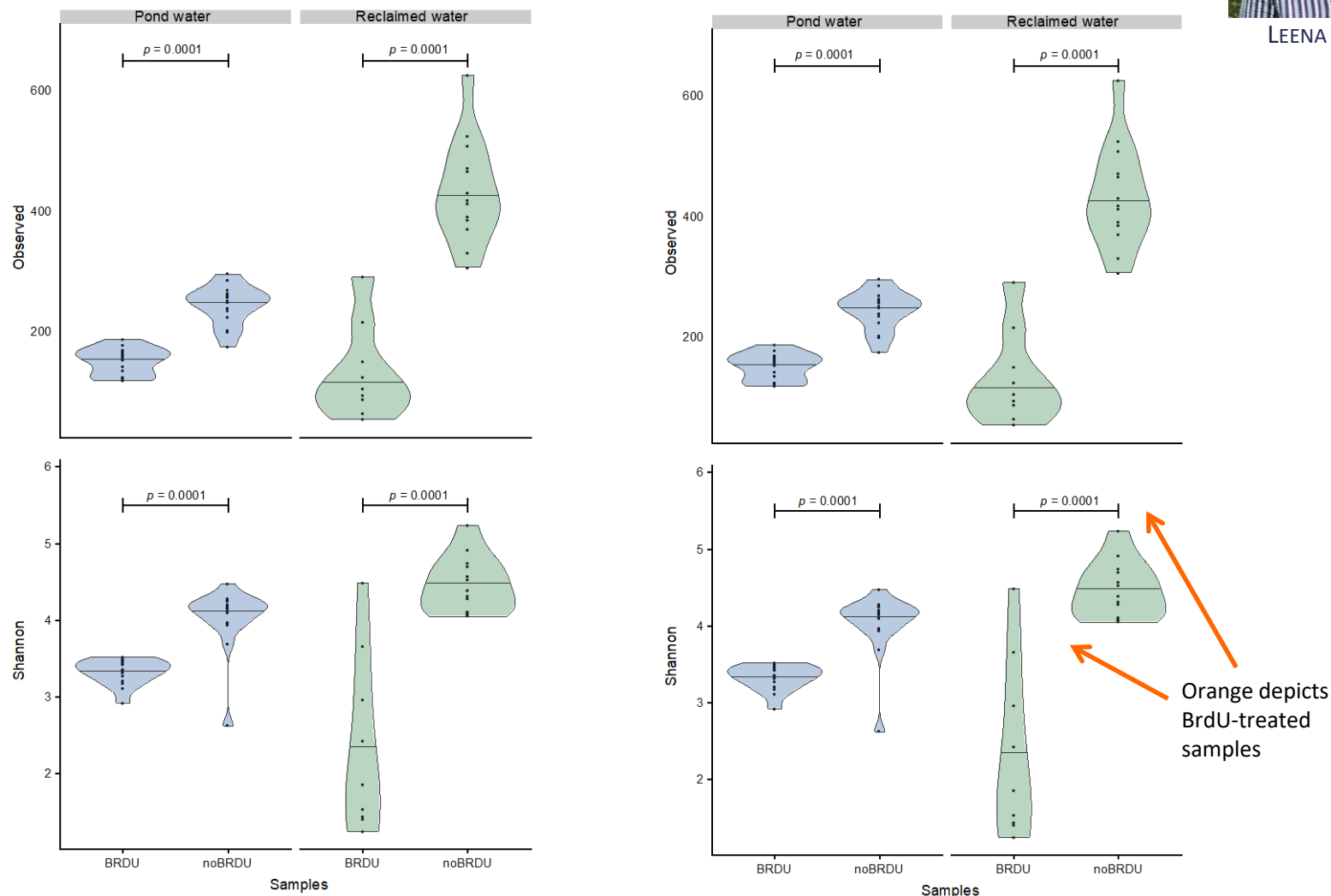


Figure 1: Alpha and beta diversity (rarefied) among BrdU- and non-BrdU-treated reclaimed water (MD06) and pond water (MD10).

Shared and Unique Bacterial Profiles in BrdU and Non-BrdU Treated Recycled Water Samples



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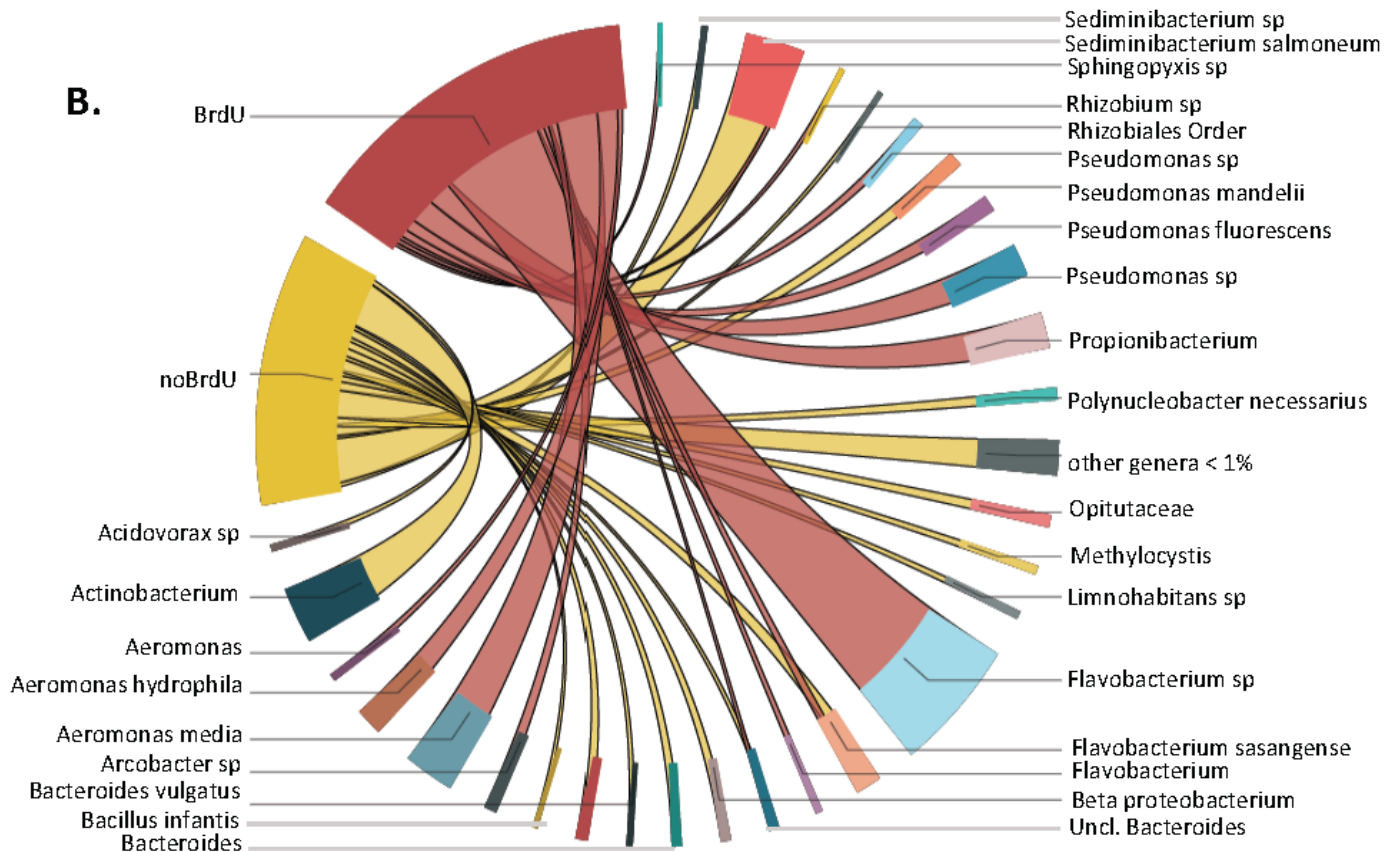


Figure 4, B: Shared and unique bacterial profiles visualized by chord plots between BrdU and non-BrdU-treated reclaimed water samples.

Are antibiotic resistance genes present in the metabolically-active fraction of bacterial communities?



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Reclaimed Water



Figure 5: Relative abundance of antibiotic resistance genes in reclaimed water (A) samples by BrdU-treatment and by sampling month.

TAKE HOME MESSAGES

- Agricultural water reuse is an important part of future water and food security solutions
- Microbiological contaminants can persist in recycled water and untreated surface water
- Larger volume sampling ($\geq 10\text{L}$) is necessary for improved pathogen detection
- Coupled DNA-labeling and sequencing approaches can help improve understanding of the microbiological risks associated with agricultural water reuse





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PUBLIC HEALTH



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The CONSERVE Team



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